

# *References*

## REFERENCES

- Baudino, T. A., (2015), “Targeted Cancer Therapy: The Next Generation of Cancer Treatment.” *Curr. Drug Discov. Technol.*, 12(1), pp. 3-20.
  - Adeyinka, A., Nui, Y., Cherlet, T., Snell, L., Watson, P. H., and Murphy, L. C., (2002), “Activated mitogen-activated protein kinase expression during human breast tumorigenesis and breast cancer progression,” *Clin. Cancer Res.*, 8(6), pp. 1747-1753.
  - Ahmad, I., Jadhav, H., Shinde, Y., Jagtap, V., Girase, R., and Patel, H., (2021), “Optimizing Bedaquiline for Cardiotoxicity by Structure Based Virtual Screening, DFT Analysis and Molecular Dynamic Simulation Studies to Identify Selective MDR-TB Inhibitors.” *In Silico Pharmacol.*, 9(1), pp. 1-15.
  - Akram, M., Iqbal, M., Daniyal, M., and Khan, A. U., (2017), “Awareness and Current Knowledge of Breast Cancer.” *Biol. Res.*, 50(1), pp. 1-23.
  - Alder, B. J., and Wainwright T. E., (1957), “Phase Transition for a Hard Sphere System,” *J. Chem. Phys.*, 27(5), pp. 1208-1209.
  - Alder, B. J., and Wainwright T. E., (1959), “Studies in Molecular Dynamics. I. General Method,” *J. Chem. Phys.*, 31(2), pp. 459-466.
  - Alessi, D. R., Cuenda, A., Cohen, P., Dudley, D. T., and Saltiel, A. R., (1995), “PD 098059 is a Specific Inhibitor of the Activation of Mitogen-Activated Protein Kinase Kinase In Vitro and In Vivo,” *J. Biol. Chem.*, 270(46), pp. 27489-27494.
  - Aly, A. A., El-Sheref, E. M., Bakheet, M. E., Mourad, M. A., Bräse, S., Ibrahim, M. A., Nieger, M., Garvalov, B. K., Dalby, K. N., and Kaoud, T. S., (2019), “Design, Synthesis and Biological Evaluation of Fused Naphthofuro [3, 2-c] Quinoline-6, 7, 12-Triones and Pyrano [3, 2-c] Quinoline-6, 7, 8, 13-Tetraones Derivatives as ERK Inhibitors with Efficacy in BRAF-Mutant Melanoma,” *Bioorg. Chem.*, 82, pp. 290-305.
  - Aly, A. A., El-Sheref, E. M., Bakheet, M. E., Mourad, M. A., Brown, A. B., Bräse, S., Nieger, M., and Ibrahim, M. A., (2018), “Synthesis of Novel 1, 2-

## REFERENCES

---

- Bis-Quinolinyl-1, 4-Naphthoquinones: ERK2 Inhibition, Cytotoxicity and Molecular Docking Studies," *Bioorg. Chem.*, 81, pp. 700-712.
- Amin, L. H., Shawer, T. Z., El-Naggar, A.M., and El-Sehrawi, H. M., (2019), "Design, Synthesis, Anticancer Evaluation and Docking Studies of New Pyrimidine Derivatives as Potent Thymidylate Synthase Inhibitors," *Bioorg. Chem.*, 91, pp. 103159.
- Asati, V., Thakur, S. S., Upmanyu, N., and Bharti, S. K., (2018), "Virtual Screening, Molecular Docking, and DFT Studies of Some Thiazolidine-2, 4-diones as Potential PIM-1 Kinase Inhibitors," *Chemistry Select.*, 3(1), pp. 127-135.
- Awadallah, F. M., Abou-Seri, S. M., Abdulla, M.M., and Georgey, H. H., (2015), "Design and Synthesis of Potent 1, 2, 4-Trisubstituted Imidazolinone Derivatives with Dual P38 $\alpha$ mapk and ERK1/2 Inhibitory Activity," *Eur. J. Med. Chem.*, 94, pp. 397-404.
- Bade, B. C., and Cruz C. S. D., (2020), "Lung cancer 2020: Epidemiology, Etiology, and Prevention." *Clin. Chest Med.*, 41(1), pp. 1-24.
- Balko, J. M., Cook, R. S., Vaught, D. B., Kuba, M. G., Miller, T. W., Bhola, N. E., Sanders, M. E., Granja-Ingram, N. M., Smith, J. J., Meszoely, I. M., and Salter, J., (2012), "Profiling of Residual Breast Cancers after Neoadjuvant Chemotherapy Identifies DUSP4 Deficiency as a Mechanism of Drug Resistance," *Nat. Med.* 18(7), pp. 1052-1059.
- Bannoura, S. F., Uddin, M., Nagasaka, M., Fazili, F., Al-Hallak, M. N., Philip, P. A., El-Rayes, B., and Azmi, A. S., (2021), "Targeting KRAS in Pancreatic Cancer: New Drugs on the Horizon," *Cancer Metastasis Rev.*, 40(3), pp. 819-835.
- Barrett, S. D., Bridges, A. J., Dudley, D. T., Saltiel, A. R., Fergus, J. H., Flamme, C. M., Delaney, A. M., Kaufman, M., LePage, S., Leopold, W. R., and Przybranowski, S .A., (2008), "The Discovery of the Benzhydroxamate MEK Inhibitors CI-1040 and PD 0325901," *Bioorg. Med. Chem. Lett.*,

## REFERENCES

---

- 18(24), pp. 6501-6504.
- Bendell, J. C., Javle, M., Bekaii-Saab, T. S., Finn, R.S., Wainberg, Z. A., Laheru, D. A., Weekes, C. D., Tan, B. R., Khan, G. N., Zalupski, M. M., and Infante, J. R., (2017), “A Phase 1 Dose-Escalation and Expansion Study of Binimetinib (MEK162), A Potent and Selective Oral MEK1/2 Inhibitor,” *Br. J. Cancer.*, 116(5), pp. 575-583.
  - Blackadar, C. B., (2016), “Historical Review of the Causes of Cancer,” *World J. Clin. Oncol.*, 7(1), pp. 54.
  - Blake, J. F., Gaudino, J. J., De Meese, J., Mohr, P., Chicarelli, M., Tian, H., Garrey, R., Thomas, A., Siedem, C. S., Welch, M. B., and Kolakowski, G., (2014), “Discovery of 5, 6, 7, 8-Tetrahydropyrido [3, 4-d] Pyrimidine Inhibitors of ERK2.” *Bioorg. Med. Chem. Lett.*, 24(12), pp. 2635-2639.
  - Bochevarov, A. D., Harder, E., Hughes, T. F., Greenwood, J. R., Braden, D. A., Philipp, D. M., Rinaldo, D., Halls, M. D., Zhang, J., and Friesner, R. A., (2013), “Jaguar: A High-Performance Quantum Chemistry Software Program with Strengths in Life and Materials Sciences,” *Int. J. Quantum Chem.*, 113(18), pp. 2110-2142.
  - Boga, S. B., Alhassan, A. B., Cooper, A. B., Doll, R., Shih, N. Y., Shipps, G., Deng, Y., Zhu, H., Nan, Y., Sun, R., and Zhu, L., (2018), “Discovery of 3 (S)-thiomethyl Pyrrolidine ERK Inhibitors for Oncology,” *Bioorg. Med. Chem. Lett.*, 28(11), pp. 2029-2034.
  - Boraei, A. T., Singh, P. K., Sechi, M., and Satta, S., (2019). “Discovery of Novel Functionalized 1, 2, 4-Triazoles as PARP-1 Inhibitors in Breast Cancer: Design, Synthesis and Antitumor Activity Evaluation.” *Eur. J. Med. Chem.*, 182, pp. 111621.
  - Braicu, C., Buse, M., Busuioc, C., Drula, R., Gulei, D., Raduly, L., Rusu, A., Irimie, A., Atanasov, A. G., Slaby, O., and Ionescu, C., (2019), “A Comprehensive Review on MAPK: A Promising Therapeutic Target in Cancer,” *Cancers.*, 11(10), pp. 1618.
-

## REFERENCES

---

- Chemical ComputingGroup, M. (2008). Molecular Operating Environment, 2008.10, Chemical Computing Group Quebec, QC, Canada.
- Chiacchio, M. A., Iannazzo, D., Romeo, R., Giofrè, S. V., and Legnani, L., (2019), “Pyridine and Pyrimidine Derivatives as Privileged Scaffolds in Biologically Active Agents.” *Curr. Med. Chem.*, 26(40), pp. 7166-7195.
- Cohen, J. V., and Sullivan R. J., (2019), “Developments in the Space of New MAPK Pathway Inhibitors for BRAF-mutant Melanoma,” *Clin. Cancer Res.*, 25(19), pp. 5735-5742.
- Coveney, P., Giordanetto, F., and Stringfellow, N., (2002), “Obtaining Scalable Performance from Molecular Dynamics Codes on HPC Machines,” *Proceedings From The CUG Summit*, London, UK, pp. 20-24.
- Craig, P., and Petticrew M., (2013), “Developing and Evaluating Complex Interventions: Reflections on the 2008 MRC Guidance,” *Int. J. Nurs. Stud.*, 50(5), pp. 585-587.
- Dai, X., Xiang, L., Li, T., and Bai, Z., (2016), “Cancer Hallmarks, Biomarkers and Breast Cancer Molecular Subtypes,” *J. Cancer.*, 7(10), pp. 1281.
- Denkert, C., Liedtke, C., Tutt, A., and Von Minckwitz, G., (2017), “Molecular Alterations in Triple-Negative Breast Cancer—The Road to New Treatment Strategies” *Lancet.*, 389(10087), pp. 2430-2442.
- Diao, P. C., Lin, W. Y., Jian, X. E., Li, Y. H., You, W. W., and Zhao, P. L., (2019), “Discovery of Novel Pyrimidine-Based Benzothiazole Derivatives as Potent Cyclin-Dependent Kinase 2 Inhibitors with Anticancer Activity,” *Eur. J. Med. Chem.*, 179, pp. 196-207.
- Dong, J., Wang, N. N., Yao, Z. J., Zhang, L., Cheng, Y., Ouyang, D., Lu, A. P., and Cao, D. S., (2018), “ADMET lab: A Platform for Systematic ADMET Evaluation Based on a Comprehensively Collected ADMET Database.” *J. Cheminformatics.*, 10(1), pp. 29.
- Dong, Q., Dougan, D. R., Gong, X., Halkowycz, P., Jin, B., Kanouni, T., O’ Connell, S. M., Scorah, N., Shi, L., Wallace, M. B., and Zhou, F., (2011),

## REFERENCES

---

- “Discovery of TAK-733, a Potent and Selective MEK Allosteric Site Inhibitor for the Treatment of Cancer,” *Bioorg. Med. Chem. Lett.*, 21(5), pp. 1315-1319.
- Elimimian, E. B., Elson, L., Li, H., Liang, H., Bilani, N., Zabor, E. C., Statler, A., and Nahleh, Z., (2021), “Male Breast Cancer: A Comparative Analysis from the National Cancer Database,” *World J. Men's Health.*, 39(3), pp. 506.
  - Eralp, Y., Derin, D., Ozluk, Y., Yavuz, E., Guney, N., Saip, P., Muslumanoglu, M., Igci, A., Kücük, S., Dincer, M., and Aydiner, A., (2008), “MAPK Overexpression is Associated with Anthracycline Resistance and Increased Risk for Recurrence in Patients with Triple-Negative Breast Cancer,” *Ann. Oncol.* 19(4), pp. 669-674.
  - Farag, A. K., Hassan, A. H., Ahn, B. S., Park, K. D. and Roh, E. J., (2020), “Reprofiling of Pyrimidine-Based DAPK1/CSF1R Dual Inhibitors: Identification of 2, 5-Diamino-4-Pyrimidinol Derivatives as Novel Potential Anticancer Lead Compounds,” *J. Enzyme Inhib. Med. Chem.* 35(1), pp. 311-324.
  - Fell, J. B., Fischer, J. P., Baer, B. R., Blake, J. F., Bouhana, K., Briere, D. M., Brown, K. D., Burgess, L. E., Burns, A. C., Burkard, M. R., and Chiang, H., (2020), “Identification of the Clinical Development Candidate MRTX849, A Covalent KRASG12C Inhibitor for the Treatment of Cancer,” *J. Med. Chem.*, 63(13), pp. 6679-6693.
  - Fiser, A. and Do R. K. G., (2000), “Modeling of Loops in Protein Structures.” *Protein Sci.*, 9(9), pp. 1753-1773.
  - Forli, W., Halliday, S., Belew, R., and Olson, A. J., (2012), “AutoDock Version 4.2,” *J. Med. Chem.*, 55, 623-638.
  - Friesner, R. A., Murphy, R. B., Repasky, M. P., Frye, L. L., Greenwood, J. R., Halgren, T. A., Sanschagrin, P. C., and Mainz, D. T., (2006), “Extra Precision Glide: Docking and Scoring Incorporating a Model of Hydrophobic Enclosure for Protein–Ligand Complexes,” *J. Med. Chem.*, 49(21), pp. 6177-6196.
-

## REFERENCES

---

- Gao, J. L., Lv, G. Y., He, B. C., Zhang, B. Q., Zhang, H., Wang, N., Wang, C. Z., Du, W., Yuan, C. S., and He, T. C., (2013), “Ginseng Saponin Metabolite 20 (S)-Protopanaxadiol Inhibits Tumor Growth by Targeting Multiple Cancer Signaling Pathways,” *Oncol. Rep.*, 30(1), pp. 292-298.
- Garcia-Estevez, L., and Moreno-Bueno G., (2019), “Updating the Role of Obesity and Cholesterol in Breast Cancer,” *Breast Cancer Res.*, 21(1), pp. 1-8.
- Garrington, T. P., and Johnson G. L., (1999), “Organization and Regulation of Mitogen-Activated Protein Kinase Signaling Pathways,” *Curr. Opin. Cell Biol.*, 11(2), pp. 211-218.
- Genheden, S., and Ryde U., (2015), “The MM/PBSA and MM/GBSA Methods to Estimate Ligand-Binding Affinities,” *Expert Opin. Drug Discov.*, 10(5), pp. 449-461.
- Gill, P. M., Johnson, B. G., Pople, J. A., and Frisch, M. J., (1992), “The Performance of the Becke -Lee-Yang-Parr (B-LYP) Density Functional Theory with Various Basis Sets,” *Chem. Phys. Lett.* 197(4-5), pp. 499-505.
- Giltnane, J. M., and Balko J. M., (2014), “Rationale for Targeting the Ras/MAPK Pathway in Triple-Negative Breast Cancer,” *Discov. Med.*, 17(95), pp. 275-283.
- Goebel, L., Müller, M. P., Goody, R.S., and Rauh, D., (2020), “KRasG12C Inhibitors in Clinical Trials: A Short Historical Perspective,” *RSC Med. Chem.*, 11(7), pp. 760-770.
- Gong, J., Cai, C., Liu, X., Ku, X., Jiang, H., Gao, D., and Li, H., (2013), “ChemMapper: A Versatile Web Server for Exploring Pharmacology and Chemical Structure Association Based on Molecular 3D Similarity Method,” *Bioinformatics.*, 29(14), pp. 1827-1829.
- Goodford, P. J., (1985). “A Computational Procedure for Determining Energetically Favorable Binding Sites on Biologically Important Macromolecules,” *J. Med. Chem.*, 28(7), pp. 849-857.
- Goulielmaki, M., Assimomitis, N., Rozanc, J., Taki, E., Christodoulou, I.,

## REFERENCES

---

- Alexopoulos, L. G., Zoumpourlis, V., Pintzas, A., and Papahatjis, D., (2019), “DPS-2: A Novel Dual MEK/ERK and PI3K/AKT Pathway Inhibitor with Powerful Ex Vivo and In Vivo Anticancer Properties,” *Transl. Oncol.*, 12(7), pp. 932-950.
- Govindan, R., Fakih, M. G., Price, T. J., Falchook, G. S., Desai, J., Kuo, J. C., Strickler, J. H., Krauss, J. C., Li, B. T., Denlinger, C. S., and Durm, G., (2019), “Phase I study of AMG 510, A Novel Molecule Targeting KRAS G12C Mutant Solid Tumours,” *Ann. Oncol.*, 30, pp. v163-v164.
- Grimaldi, A. M., Simeone, E., and Ascierto, P.A., (2014), “The Role of MEK Inhibitors in the Treatment of Metastatic Melanoma,” *Curr. Opin. Oncol.*, 26(2), pp. 196-203.
- Halawa, A. H., Eskandrani, A. A., Elgammal, W. E., Hassan, S. M., Hassan, A. H., Ebrahim, H. Y., Mehany, A., El-Agrody, A.M., and Okasha, R. M., (2019), “Rational Design and Synthesis of Diverse Pyrimidine Molecules Bearing Sulfonamide Moiety as Novel ERK Inhibitors,” *Int. J. Mol. Sci.*, 20(22), pp. 5592.
- Halgren, T. A., Murphy, R. B., Friesner, R. A., Beard, H. S., Frye, L. L., Pollard, W. T., and Banks, J. L., (2004), “Glide: A New Approach for Rapid, Accurate Docking and Scoring. 2. Enrichment Factors in Database Screening,” *J. Med. Chem.*, 47(7), pp. 1750-1759.
- Hammes, G. G., (2002), “Multiple conformational changes in enzyme catalysis,” *Biochemistry.*, 41(26), pp. 8221-8228.
- Hanahan, D., and Weinberg R. A., (2011), “Hallmarks of Cancer: The Next Generation.” *Cell.*, 144(5), pp. 646-674.
- Hao, G.F., Jiang, W., Ye, Y. N., Wu, F. X., Zhu, X. L., Guo, F. B., and Yang, G.F., (2016), “ACFIS: A Web Server for Fragment-Based Drug Discovery,” *Nucleic Acids Res.*, 44(W1), pp. W550-W556.
- Harder, E., Damm, W., Maple, J., Wu, C., Reboul, M., Xiang, J. Y., Wang, L., Lupyán, D., Dahlgren, M. K., Knight, J. L. and Kaus, J. W., (2016), “OPLS3:

## REFERENCES

---

- A Force Field Providing Broad Coverage of Drug-Like Small Molecules and Proteins,” J. Chem. Theory Comput., 12(1), pp. 281-296.
- Hatzivassiliou, G., Liu, B., Brien, C. O., Spoerke, J. M., Hoeflich, K. P., Haverty, P. M., Soriano, R., Forrest, W. F., Heldens, S., Chen, H., and Toy, K., (2012), “ERK Inhibition Overcomes Acquired Resistance to MEK Inhibitors,” Mol. Cancer Ther., 11(5), pp. 1143-1154.
- Heightman, T. D., Berdini, V., Braithwaite, H., Buck, I. M., Cassidy, M., Castro, J., Courtin, A., Day, J. E., East, C., Fazal, L. and Graham, B., (2018), “Fragment-Based Discovery of a Potent, Orally Bioavailable Inhibitor that Modulates the Phosphorylation and Catalytic Activity of ERK1/2,” J. Med. Chem., 61(11), pp. 4978-4992.
- Hoeflich, K. P., Brien, C. O., Boyd, Z., Cavet, G., Guerrero, S., Jung, K., Januario, T., Savage, H., Punnoose, E., Truong, T., and Zhou, W., (2009), “In Vivo Antitumor Activity of MEK and Phosphatidylinositol 3-Kinase Inhibitors in Basal-Like Breast Cancer Models,” Clin. Cancer Res., 15(14), pp. 4649-4664.
- Housman, G., Byler, S., Heerboth, S., Lapinska, K., Longacre, M., Snyder, N., and Sarkar, S., (2014), “Drug Resistance in Cancer: An Overview” Cancers., 6(3), pp. 1769-1792.
- Hu, Y., Stumpfe, D., and Bajorath, J., (2017), “Recent Advances in Scaffold Hopping: Miniperspective,” J. Med. Chem., 60(4), pp. 1238-1246.
- Hubbard, R. E., Jahnke, W., and Erlanson, D.A., (2016), “Fragment-based Drug Discovery Lessons and Outlook,” Wiley-VCH, Weinheim Germany, pp. 3-29.
- Hyun, S., and Shin D., (2021), “Small-Molecule Inhibitors and Degraders Targeting KRAS-Driven Cancers,” Int. J. Mol. Sci., 22(22), pp. 12142.
- Inamdar, G. S., Madhunapantula, S. V., and Robertson, G.P., (2010). “Targeting the MAPK Pathway in Melanoma: Why some Approaches Succeed and other Fail.” Biochem. Pharmacol., 80(5), pp. 624-637.

## REFERENCES

---

- Jacobson, M. P., Pincus, D. L., Rapp, C. S., Day, T. J., Honig, B., Shaw, D. E. and Friesner, R. A., (2004), “A Hierarchical Approach to All-Atom Protein Loop Prediction,” *Proteins.*, 55(2), pp. 351-367.
- Jang, S., and Atkins M., (2014), “Treatment of BRAF-mutant Melanoma: The Role of Vemurafenib and other Therapies,” *Clin. Pharmacol. Ther.*, 95(1), pp. 24-31.
- Jia, Y., Quinn, C. M., Kwak, S. and Talanian, R. V., (2008), “Current In Vitro Kinase Assay Technologies: The Quest for a Universal Format,” *Curr. Drug Discov. Technol.*, 5(1), pp. 59-69.
- Jiang, W., Wang, X., *et al.*, (2020), “Expression and Clinical Significance of MAPK and EGFR in Triple- Negative Breast Cancer,” *Oncology Letters*, 19(3), pp. 1842-1848.
- Jin, X. Y., Chen, H., Zhang, C., Xue, L., and Yang, L., (2019), “Design, Synthesis, and Anticancer Evaluation of Novel Quinoline Derivatives of Ursolic Acid with Hydrazide, Oxadiazole, and Thiadiazole Moieties as Potent MEK Inhibitors,” *J. Enzyme Inhib. Med. Chem.*, 34(1), pp. 955-972.
- Joshi, G., Nayyar, H., Marin Alex, J., Vishwakarma, S. G., Mittal, S., and Kumar, R., (2016), “Pyrimidine-fused Derivatives: Synthetic Strategies and Medicinal Attributes,” *Curr. Top Med. Chem.*, 16(28), pp. 3175-3210.
- Kapetanovic, I., (2008), “Computer-Aided Drug Discovery and Development (CADD): In Silico-Chemico-Biological Approach,” *Chem. Biol. Interact.*, 171(2), pp. 165-176.
- Ke, X., and Shen L., (2017), “Molecular Targeted Therapy of Cancer: The Progress and Future Prospect,” *Front. Lab. Med.*, 1(2), pp. 69-75.
- Kemp, W., (1991), “Nuclear Magnetic Resonance Spectroscopy,” *Organic Spectroscopy*, Springer, Palgrave, London, pp. 101-241.
- Kidger, A. M., Sipthorp, J., and Cook, S. J., (2018), “ERK1/2 inhibitors: New Weapons to Inhibit the RAS-regulated RAF-MEK1/2-ERK1/2 Pathway,” *Pharmacol. Ther.*, 187, pp. 45-60.

## REFERENCES

---

- Kim, D., Kim, S. Y., Kim, D., Yoon, N. G., Yun, J., Hong, K. B., Lee, C., Lee, J. H., Kang, B. H., and Kang, S., (2020), “Development of Pyrazolo [3, 4-*d*] Pyrimidine-6-amine-based TRAP1 Inhibitors that Demonstrate In Vivo Anticancer Activity in Mouse Xenograft Models,” *Bioorg. Chem.*, 101, pp. 103901.
- Kim, E. K., and Choi E. J., (2010). “Pathological Roles of MAPK Signaling Pathways in Human Diseases,” *Biochim. Biophys. Acta. Mol. Basis Dis.*, 1802(4), pp. 396-405.
- Kolb, P., and Caflisch A., (2006), “Automatic and Efficient Decomposition of Two-Dimensional Structures of Small Molecules for Fragment-Based High-Throughput Docking,” *J. Med. Chem.*, 49(25), pp. 7384-7392.
- Korzeniecki, C., and Priefer R., (2021), “Targeting KRAS Mutant Cancers by Preventing Signaling Transduction in the MAPK Pathway,” *Eur. J. Med. Chem.*, 211, pp. 113006.
- Kumar, B., Sharma, P., Gupta, V. P., Khullar, M., Singh, S., Dogra, N. and Kumar, V., (2018), “Synthesis and Biological Evaluation of Pyrimidine Bridged Combretastatin Derivatives as Potential Anticancer Agents and Mechanistic Studies,” *Bioorg. Chem.*, 78, pp. 130-140.
- Kumar, S., and Narasimhan B., (2018), “Therapeutic Potential of Heterocyclic Pyrimidine Scaffolds,” *Chem. Cent. J.*, 12(1), pp. 1-29.
- Kumar S., P., and Silakari O., (2019), “In Silico Guided Development of Imine-Based Inhibitors for Resistance-Deriving Kinases,” *J. Biomol. Struct. Dyn.*, 37(10), pp. 2593-2599.
- Kushwah, V., Agrawal, A. K., Dora, C. P., Mallinson, D., Lamprou, D. A., Gupta, R. C., and Jain, S., (2017), “Novel Gemcitabine Conjugated Albumin Nanoparticles: a Potential Strategy to Enhance Drug Efficacy in Pancreatic Cancer Treatment,” *Pharm. Res.*, 34(11), pp. 2295-2311.
- Kushwah, V., Katiyar, S. S., Dora, C. P., Agrawal, A. K., Lamprou, D. A., Gupta, R. C., and Jain, S., (2018), “Co-delivery of Docetaxel and Gemcitabine

## REFERENCES

---

- by Anacardic Acid Modified Self-Assembled Albumin Nanoparticles for Effective Breast Cancer Management," *Acta Biomater.*, 73, pp. 424-436.
- Lau, W. S., Chen, W. F., Chan, R. Y. K., Guo, D. A., and Wong, M. S., (2009), "Mitogen-Activated Protein Kinase (MAPK) Pathway Mediates the Oestrogen-Like Activities of Ginsenoside Rg1 in Human Breast Cancer (MCF-7) Cells," *Br. J. Pharmacol.*, 156(7), pp. 1136-1146.
- Lee, C., Yang, W., and Parr, R. G., (1988), "Development of the Colle-Salvetti Correlation-Energy Formula into a Functional of the Electron Density," *Phys. Rev. B.*, 37(2), pp. 785.
- Lee, S., Rauch, J., and Kolch, W., (2020), "Targeting MAPK Signaling in Cancer: Mechanisms of Drug Resistance and Sensitivity," *Int. J. Mol. Sci.*, 21(3), pp. 1102.
- Lee, Y. T., Tan, Y. J., and Oon, C. E., (2018), "Molecular Targeted Therapy: Treating Cancer with Specificity," *Eur. J. Pharmacol.*, 834, pp. 188-196.
- Leijen, S., Middleton, M. R., Tresca, P., Kraeber-Bodéré, F., Dieras, V., Scheulen, M. E., Gupta, A., Lopez-Valverde, V., Xu, Z. X., Rueger, R., and Tessier, J. J., (2012), "Phase I Dose-Escalation Study of the Safety, Pharmacokinetics, and Pharmacodynamics of the MEK Inhibitor RO4987655 (CH4987655) in Patients with Advanced Solid Tumors," *Clin. Cancer Res.*, 18(17), pp. 4794-4805.
- Li, J. X., Feng, J. M., Wang, Y., Li, X. H., Chen, X. X., Su, Y., Shen, Y. Y., Chen, Y., Xiong, B., Yang, C. H., and Ding, J., (2014), "The B-Raf V600E Inhibitor Dabrafenib Selectively Inhibits RIP3 and Alleviates Acetaminophen-Induced Liver Injury." *Cell Death Dis.*, 5(6), pp. e1278-e1278.
- Li, L., Hou, X., Xu, R., Liu, C., and Tu, M., (2017), "Research Review on the Pharmacological Effects of Astragaloside IV," *Fundam. Clin. Pharmacol.* 31(1), pp. 17-36.
- Li, L., Liu, F., Jin, N., Tang, S., Chen, Z., Yang, X., Ding, J., Geng, M., Jiang, L., Huang, M., and Cao, J., (2016), "Discovery and Structure Activity

## REFERENCES

---

- Relationship Study of Novel Indazole Amide Inhibitors for Extracellular Signal-Regulated Kinase1/2 (ERK1/2)," Bioorg. Med. Chem. Lett. 26(11), pp. 2600-2604.
- Liu, F., Yang, X., Geng, M., and Huang, M., (2018), "Targeting ERK, an Achilles' Heel of the MAPK Pathway, in Cancer Therapy," Acta Pharm. Sin. 8(4), pp. 552-562.
  - Liu, K., Rao, W., Parikh, H., Li, Q., Guo, T. L., Grant, S., Kellogg, G. E., and Zhang, S., (2012), "3, 5-Disubstituted-thiazolidine-2, 4-dione Analogs as Anticancer Agents: Design, Synthesis and Biological Characterization," Eur. J. Med. Chem., 47, pp. 125-137.
  - Luebker, S. A., and Koepsell S. A., (2019), "Diverse Mechanisms of BRAF Inhibitor Resistance in Melanoma Identified in Clinical and Preclinical Studies." Front. Oncol., 9, pp. 268.
  - Ma, H., Deacon, S., and Horiuchi, K., (2008), "The Challenge of Selecting Protein Kinase Assays for Lead Discovery Optimization," Expert Opin. Drug Discov., 3(6), pp. 607-621.
  - Macalino, S. J. Y., Gosu, V., Hong, S., and Choi, S., (2015), "Role of Computer-Aided Drug Design in Modern Drug Discovery," Arch. Pharm. Res., 38(9), pp. 1686-1701.
  - Mangana, J., Levesque, M. P., Karpova, M. B., and Dummer, R., (2012), "Sorafenib in Melanoma." Expert Opin. Investig. Drugs., 21(4), pp. 557-568.
  - Markham, A., and Keam S. J., (2020), "Selumetinib: First Approval." Drugs., 80, pp. 931-937.
  - Martin-Liberal, J., and Larkin J., (2014), "New RAF Kinase Inhibitors in Cancer Therapy," Expert Opin. Investig. Drugs., 15(9), pp. 1235-1245.
  - Martinelli, E., Troiani, T., Aiuto, D. E., Morgillo, F., Vitagliano, D., Capasso, A., Costantino, S., Ciuffreda, L.P., Merolla, F., Vecchione, L., and De Vriendt, V., (2013), "Antitumor Activity of Pimasertib, a Selective MEK 1/2 Inhibitor, in Combination with PI3K/Mtor Inhibitors or with Multi-Targeted

## REFERENCES

---

- Kinase Inhibitors in Pimasertib-Resistant Human Lung And Colorectal Cancer Cells," Int. J. Cancer., 133(9), pp. 2089-2101.
- Marzo, I., and Naval J., (2013), "Antimitotic Drugs in Cancer Chemotherapy: Promises and Pitfalls," Biochem. Pharmacol., 86(6), pp. 703-710.
  - Masliah-Planchon, J., Garinet, S., and Pasman, E., (2016), "RAS-MAPK Pathway Epigenetic Activation in Cancer: Mirnas in Action," Oncotarget., 7(25), pp. 38892.
  - McCommon, J. A., Gelin, B. R., and Karplus, M., (1977), "Dynamics of Folded Proteins." Nature., 267(5612), pp. 585-590.
  - McCubrey, J. A., Steelman, L. S., Kempf, C. R., Chappell, W. H., Abrams, S. L., Stivala, F., Malaponte, G., Nicoletti, F., Libra, M., Baesecke, J., and Maksimovic-Ivanic, D., (2011), "Therapeutic Resistance Resulting from Mutations in Raf/MEK/ERK and PI3K/PTEN/Akt/Mtor Signaling Pathways," J. Cell. Physiol., 226(11), pp. 2762-2781.
  - Metwally, N. H., and Deeb E. A., (2018), "Synthesis, Anticancer Assessment on Human Breast, Liver and Colon Carcinoma Cell Lines and Molecular Modeling Study using Novel Pyrazolo [4, 3-c] Pyridine Derivatives," Bioorg. Chem., 77, pp. 203-214.
  - Mohamed, M. M., Khalil, A. K., Abbass, E. M., and El-Naggar, A. M., (2017), "Design, Synthesis of New Pyrimidine Derivatives as Anticancer and Antimicrobial Agents," Synth. Commun., 47(16), pp. 1441-1457.
  - Mohana R, S., and Sompalle R., (2016), "Synthetic Chemistry of Pyrimidines and Fused Pyrimidines: A Review," Synth. Commun., 46(8), pp. 645-672.
  - Montagut, C., and Settleman J., (2009), "Targeting the RAF-MEK-ERK Pathway in Cancer Therapy," Cancer Lett., 283(2), pp. 125-134.
  - Morgan, S., Grootendorst, P., Lexchin, J., Cunningham, C. and Greyson, D., (2011), "The Cost of Drug Development: A Systematic Review," Health Policy., 100(1), pp. 4-17.
  - Mukherjee, S., Balias, T. E., and Rizzo, R. C., (2010), "Docking Validation

## REFERENCES

---

- Resources: Protein Family and Ligand Flexibility Experiments," J. Chem. Inf. Model., 50(11), 1986-2000.
- Nasser, A. A., Eissa, I. H., Oun, M. R., El-Zahabi, M. A., Taghour, M. S., Belal, A., Saleh, A. M., Mehany, A. B., Luesch, H., Mostafa, A. E. and Afifi, W. M., (2020), "Discovery of New Pyrimidine-5-Carbonitrile Derivatives as Anticancer Agents Targeting EGFR WT and EGFR T790M," Org. Biomol. Chem., 18(38), pp. 7608-7634.
  - Nissan, M. H., Pratillas, C. A., Jones, A. M., Ramirez, R., Won, H., Liu, C., Tiwari, S., Kong, L., Hanrahan, A. J., Yao, Z. and Merghoub, T., (2014), "Loss of NF1 in Cutaneous Melanoma is Associated with RAS Activation and MEK Dependence," Cancer Res., 74(8), pp. 2340-2350.
  - Normanno, N., Luca, A. D., Maiello, M. R., Campiglio, M., Napolitano, M., Mancino, M., Carotenuto, A., Viglietto, G., and Menard, S., (2006), "The MEK/MAPK Pathway is Involved in the Resistance of Breast Cancer Cells to the EGFR Tyrosine Kinase Inhibitor Gefitinib," J. Cell. Physiol., 207(2), pp. 420-427.
  - Ostrem, J. M., Peters, U., Sos, M. L., Wells, J. A. and Shokat, K. M., (2013), "K-Ras (G12C) Inhibitors Allosterically Control GTP Affinity and Effector Interactions," Nature., 503(7477), pp. 548-551.
  - Pal, S. K., Childs, B. H., and Pegram, M., (2011), "Triple Negative Breast Cancer: Unmet Medical Needs," Breast Cancer Res. Treat., 125(3), pp. 627-636.
  - Park, H. K., Jeong, H., Ko, E., Lee, G., Lee, J. E., Lee, S. K., Lee, A. J., Im, J. Y., Hu, S., Kim, S. H., and Lee, J. H., (2017), "Paralog Specificity Determines Subcellular Distribution, Action Mechanism, and Anticancer Activity of TRAP1 Inhibitors." J. Med. Chem., 60(17), pp. 7569-7578.
  - Pathania, S., and Rawal R. K., (2020), "An Update on Chemical Classes Targeting ERK1/2 for the Management of Cancer," Future Med. Chem., 12(7), pp. 593-611.

## REFERENCES

---

- Pavia, D. L., Lampman, G. M., Kriz, G. S. and Vyvyan, J. A., (2014), Introduction to Spectroscopy, Cengage Learning, Bellingham, Washington.
- Peng, B., He, R., Xu, Q., Yang, Y., Hu, Q., Hou, H., Liu, X., and Li, J., (2019), "Ginsenoside 20 (S)-Protopanaxadiol Inhibits Triple-Negative Breast Cancer Metastasis In Vivo by Targeting EGFR-Mediated MAPK Pathway," *Pharmacol. Res.*, 142, pp. 1-13.
- Pouysségur, J., and Lenormand P., (2016), "ERK1 and ERK2 Map Kinases: Specific Roles or Functional Redundancy?," *Front. Cell Dev. Biol.*, 4, pp. 53.
- Pratillas, C. A., Taylor, B. S., Ye, Q., Viale, A., Sander, C., Solit, D. B., and Rosen, N., (2009), "V600EBRAF is Associated with Disabled Feedback Inhibition of RAF-MEK Signaling and Elevated Transcriptional Output of the Pathway," *Proc. Natl. Acad. Sci.*, 106(11), pp. 4519-4524.
- Queiroz, A. N., Gomes, B. A., Moraes Jr, W. M., and Borges, R. S., (2009), "A Theoretical Antioxidant Pharmacophore for Resveratrol." *Eur. J. Med. Chem.*, 44(4), pp. 1644-1649.
- Rahman, A., (1964), "Correlations in the Motion of Atoms in Liquid Argon." *Phys. Rev.*, 136(2A), pp. A405.
- Rawal, R. K., Prabhakar, Y. S., Katti, S. B., and Clercq, D. E., (2005), "2-(Aryl)-3-furan-2-ylmethyl-thiazolidin-4-ones as Selective HIV-RT Inhibitors," *Bioorg. Med. Chem.*, 13(24), pp. 6771-6776.
- Rawal, R. K., Tripathi, R., Katti, S. B., Pannecouque, C., and Clercq, D. E., (2007), "Design, Synthesis, and Evaluation of 2-aryl-3-heteroaryl-1, 3-thiazolidin-4-ones as Anti-HIV Agents," *Bioorg. Med. Chem.*, 15(4), pp. 1725-1731.
- Rawal, R. K., Tripathi, R., Katti, S. B., Pannecouque, C., and Clercq, D. E., (2008), "Design and Synthesis of 2-(2, 6-dibromophenyl)-3-heteroaryl-1, 3-thiazolidin-4-ones as Anti-HIV Agents." *Eur. J. Med. Chem.* 43(12), pp. 2800-2806.
- Reddy, D. S., Kongot, M., Singh, V., Siddiquee, M. A., Patel, R., Singhal, N.

## REFERENCES

---

- K., Avecilla, F., and Kumar, A., (2021), “Biscoumarin–Pyrimidine Conjugates as Potent Anticancer Agents and Binding Mechanism of Hit Candidate with Human Serum Albumin,” *Arch. Pharm.*, 354(1), pp. 2000181.
- Rees, D. C., Congreve, M., Murray, C. W. and Carr, R., (2004), “Fragment-Based Lead Discovery.” *Nat. Rev. Drug Discov.*, 3(8), pp. 660-672.
- Ren, L., Grina, J., Moreno, D., Blake, J. F., Gaudino, J. J., Garrey, R., Metcalf, A. T., Burkard, M., Martinson, M., Rasor, K. and Chen, H., (2015), “Discovery of Highly Potent, Selective, and Efficacious Small Molecule Inhibitors of ERK1/2,” *J. Med. Chem.*, 58(4), pp. 1976-1991.
- Rice, K. D., Aay, N., Anand, N. K., Blazey, C. M., Bowles, O. J., Bussenius, J., Costanzo, S., Curtis, J. K., Defina, S. C., Dubenko, L. and Engst, S., (2012), “Novel Carboxamide-Based Allosteric MEK Inhibitors: Discovery and Optimization Efforts Toward XL518 (GDC-0973).” *ACS Med. Chem. Lett.*, 3(5), pp. 416-421.
- Roskoski Jr, R., (2021), “Blockade of Mutant RAS Oncogenic Signaling with a Special Emphasis on KRAS,” *Pharmacol. Res.*, 172, pp. 105806.
- Valdeira, A. S., Ritt, D. A., Morrison, D. K., McMahon, J. B., Gustafson, K. R., and Salvador, J. A., (2018), “Synthesis and Biological Evaluation of New Madecassic Acid Derivatives Targeting ERK Cascade Signaling.” *Front. Chem.*, 6, pp. 434.
- Samatar, A. A., (2017), “Extracellular Signal-Regulated Kinase (ERK1 and ERK2) Inhibitors,” *Conquering RAS.*, Academic Press, Cambridge, US, pp. 233-249, Chap. 13.
- Sammons, R. M., Perry, N. A., Li, Y., Cho, E. J., Piserchio, A., Zamora-Olivares, D. P., Ghose, R., Kaoud, T. S., Debevec, G., Bartholomeusz, C., Gurevich V. V., Iverson T. M., Giulianotti, M., Houghten R. A., and Dalby K. N., (2019), “A Novel Class of Common Docking Domain Inhibitors That Prevent ERK2 Activation and Substrate Phosphorylation,” *ACS Chem. Biol.*, 14(6), pp. 1183-1194.

## REFERENCES

---

- Schmieder, R., Puehler, F., Neuhaus, R., Kissel, M., Adjei, A. A., Miner, J. N., Mumberg, D., Ziegelbauer, K., and Scholz, A., (2013), “Allosteric MEK1/2 Inhibitor Refametinib (BAY 86-9766) in Combination with Sorafenib Exhibits Antitumor Activity in Preclinical Murine and Rat Models of Hepatocellular Carcinoma.” *Neoplasia.*, 15(10), pp. 1161-1171.
- Schubbert, S., Shannon, K., and Bollag, G., (2007), “Hyperactive Ras in Developmental Disorders and Cancer.” *Nat. Rev. Cancer.*, 7(4), pp. 295-308.
- Seerden, J. P. G., Leusink-Ionescu, G., Woudenberg-Vrenken, T., Dros, B., Molema, G., Kamps, J. A., and Kellogg, R. M., (2014), “Synthesis and Structure–Activity Relationships of 4-fluorophenyl-imidazole P38 $\alpha$  MAPK, CK1 $\delta$  and JAK2 Kinase Inhibitors.” *Bioorg. Med. Chem. Lett.*, 24(15), pp. 3412-3418.
- Seger, R., and Krebs E. G., (1995), “The MAPK Signaling Cascade.” *FASEB J.*, 9(9), pp. 726-735.
- Sharma, A., Shah S. R., Illum, H., and Dowell, J., (2012), “Vemurafenib.” *Drugs*, 72(17), pp. 2207-2222.
- Shaw, D. E., (2005), “A Fast, Scalable Method for the Parallel Evaluation of Distance-Limited Pairwise Particle Interactions.” *J. Comput. Chem.*, 26(13), pp. 1318-1328.
- Siegel, R. L., Miller, K. D., Fuchs, H. E., and Jemal, A., (2021), “Cancer Statistics, 2021.” *CA- Cancer J. Clin.*, 71(1), pp. 7-33.
- Silakari, O., and Singh P. K., (2020), Concepts and Experimental Protocols of Modelling and Informatics in Drug Design, Academic Press, London, UK, Chap. 6.
- Silverstein, R. M., and Bassler G. C., (1962), “Spectrometric Identification of Organic Compounds.” *J. Chem. Educ.*, 39(11), pp. 546.
- Singh, P., and Silakari O., (2017). “Molecular Dynamics and Pharmacophore Modelling Studies of Different Subtype (ALK And EGFR (T790M)) Inhibitors in NSCLC.” *SAR QSAR Environ Res.*, 28(3), pp. 221-233.

## REFERENCES

---

- Singh, P. K., Chaudhari, D., Jain, S., and Silakari, O., (2019), “Structure Based Designing of Triazolopyrimidone-Based Reversible Inhibitors for Kinases Involved in NSCLC.” *Bioorg. Med. Chem. Lett.*, 29(13), pp. 1565-1571.
- Singh, P. K., and Silakari O., (2018), “Pharmacophore and Molecular Dynamics Based Activity Profiling of Natural Products for Kinases Involved in Lung Cancer,” *J. Mol. Model.*, 24(11), pp. 318.
- Stillinger, F. H., and Rahman A., (1974). “Improved Simulation Of Liquid Water by Molecular Dynamics,” *J. Chem. Phys.*, 60(4), pp. 1545-1557.
- Sullivan, R. J., Infante, J. R., Janku, F., Wong, D. J. L., Sosman, J. A., Keedy, V., Patel, M. R., Shapiro, G. I., Mier, J. W., Tolcher, A. W., and Wang-Gillam, A., (2018), “First-in-Class ERK1/2 Inhibitor Ulixertinib (BVD-523) in Patients with MAPK Mutant Advanced Solid Tumors: Results of a Phase I Dose-Escalation and Expansion Study.” *Cancer Discov.* 8(2), pp. 184-195.
- Van Dort, M. E., Galbán, S., Wang, H., Sebolt-Leopold, J., Whitehead, C., Hong, H., Rehemtulla, A. and Ross, B. D., (2015), “Dual Inhibition of Allosteric Mitogen-Activated Protein Kinase (MEK) And Phosphatidylinositol 3-Kinase (PI3K) Oncogenic Targets with a Bifunctional Inhibitor,” *Bioorg. Med. Chem.*, 23(7), pp. 1386-1394.
- Van Dort, M. E., Hong, H., Wang, H., Nino, C. A., Lombardi, R. L., Blanks, A. E., Galbán, S. and Ross, B. D., (2016), “Discovery of Bifunctional Oncogenic Target Inhibitors Against Allosteric Mitogen-Activated Protein Kinase (MEK1) and Phosphatidylinositol 3-Kinase (PI3K),” *J. Med. Chem.*, 59(6), pp. 2512-2522.
- Von Kriegsheim, A., Baiocchi, D., Birtwistle, M., Sumpton, D., Bienvenut, W., Morrice, N., Yamada, K., Lamond, A., Kalna, G., Orton, R., and Gilbert, D., (2009), “Cell Fate Decisions are Specified by the Dynamic ERK Interactome,” *Nat. Cell Biol.*, 11(12), pp. 1458-1464.
- Wada, M., Horinaka, M., Yamazaki, T., Katoh, N. and Sakai, T., (2014), “The

## REFERENCES

---

- Dual RAF/MEK Inhibitor CH5126766/RO5126766 may be a Potential Therapy for RAS-Mutated Tumor Cells,” PLoS One., 9(11), pp. e113217.
- Wang, L., Zhang, Q., Zhu, G., Zhang, Z., Zhi, Y., Zhang, L., Mao, T., Zhou, X., Chen, Y., Lu, T., and Tang, W., (2017), “Design, Synthesis and Evaluation of Derivatives Based on Pyrimidine Scaffold as Potent Pan-Raf Inhibitors to Overcome Resistance,” Eur. J. Med. Chem., 130, pp. 86-106.
  - Wang, Z., Wang, X., Li, Y., Lei, T., Wang, E., Li, D., Kang, Y., Zhu, F., and Hou, T., (2019), “Farppi: A Webserver for Accurate Prediction of Protein-Ligand Binding Structures for Small-Molecule PPI Inhibitors by MM/PB (GB) SA Methods.” Bioinformatics., 35(10), pp. 1777-1779.
  - Ward, R. A., Bethel, P., Cook, C., Davies, E., Debreczeni, J. E., Fairley, G., Feron, L., Flemington, V., Graham, M. A., Greenwood, R., and Griffin, N., (2017), “Structure-Guided Discovery of Potent and Selective Inhibitors of ERK1/2 from a Modestly Active and Promiscuous Chemical Start Point,” J. Med. Chem., 60(8), pp. 3438-3450.
  - Wilde, R. E., and Singh S., (1998), Statistical Mechanics: Fundamentals and Modern Applications, Wiley, New York.
  - Wong, D. J., Robert, L., Atefi, M. S., Lassen, A., Avarappatt, G., Cerniglia, M., Avramis, E., Tsoi, J., Foulad, D., Graeber, T. G., and Comin-Anduix, B., (2014), “Antitumor Activity of the ERK Inhibitor SCH722984 against BRAF Mutant, NRAS Mutant and Wild-Type Melanoma,” Mol. Cancer, 13(1), pp. 1-15.
  - Wright, C. J., and McCormack P. L., (2013), “Trametinib: First Global Approval,” Drugs., 73(11), pp. 1245-1254.
  - Xi, D., Niu, Y., Li, H., Noha, S. M., Temml, V., Schuster, D., Wang, C., Xu, F., and Xu, P., (2019), “Discovery of Carbazole Derivatives as Novel Allosteric MEK Inhibitors by Pharmacophore Modeling and Virtual Screening,” Eur. J. Med. Chem., 178, pp. 802-817.
  - Yerragunta, V., Patil, P., Anusha, V., KumaraSwamy, T., Suman, D. and

## REFERENCES

---

- Samhitha, T., (2013), "Pyrimidine and its Biological Activity: A Review," *Pharma Tutor.*, 1(2), pp. 39-44.
- Yu, Z., Chen, Z., Su, Q., Ye, S., Yuan, H., Kuai, M., Lv, M., Tu, Z., Yang, X., Liu, R., and Hu, G., (2019), "Dual Inhibitors of RAF-MEK-ERK and PI3K-PDK1-AKT Pathways: Design, Synthesis and Preliminary Anticancer Activity Studies of 3-Substituted-5-(Phenylamino) Indolone Derivatives," *Bioorg. Med. Chem.*, 27(6), pp. 944-954.
- Yuan, S., Norgard, R. J., and Stanger, B. Z., (2019), "Cellular Plasticity in Cancer," *Cancer Discov.*, 9(7), pp. 837-851.
- Zhang, Y. J., Xu, Z. G., Li, S. Q., He, L. J., Tang, Y., Chen, Z. Z., and Yang, D. L., (2018), "Benzimidazoisoquinoline Derivatives Inhibit Glioblastoma Cell Proliferation through Down-Regulating Raf/MEK/ERK and PI3K/AKT Pathways," *Cancer Cell Int.*, 18(1), pp. 1-12.
- Zhao, H., (2007), "Scaffold Selection and Scaffold Hopping in Lead Generation: A Medicinal Chemistry Perspective," *Drug Discov. Today.*, 12(3-4), pp. 149-155.
- Zhou, J., Negi, A., Mirallai, S. I., Warta, R., Herold-Mende, C., Carty, M. P., Ye, X. S., and Murphy, P.V., (2019), "N-Alkyl-1, 5-dideoxy-1, 5-imino-L-fucitols as Fucosidase Inhibitors: Synthesis, Molecular Modelling and Activity Against Cancer Cell Lines," *Bioorg. Chem.*, 84, pp. 418-433.

# APPENDIX

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Review article

Pyrrolopyrimidines: An update on recent advancements in their medicinal attributes

Shelly Pathania <sup>a,b</sup>, Ravindra K. Rawal <sup>c,\*</sup>

<sup>a</sup> Department of Pharmaceutical Chemistry, Indi-Soviet Friendship College of Pharmacy (ESCP), Moga, 142001, India  
<sup>b</sup> Department of Pharmaceutical Sciences and Technology, Maharaja Ranjit Singh Punjab Technical University, Bathinda, 151001, Punjab, India  
<sup>c</sup> Department of Chemistry, Maharishi Markandeshwar (Deemed to be University), Mullana, 133207, Ambala, Haryana, India

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ABSTRACT

Fused heterocycles are reported to demonstrate variety of biological activities such as anticancer, antibacterial, antifungal and anti-inflammatory, and are thus exhaustively utilized in the field of medicinal chemistry. Pyrrolopyrimidines is one of the major classes of fused heterocycles which are extensively reported throughout the literature. Several reports suggest that pyrrolopyrimidine as fused scaffold possess more diverse and potent pharmacological profile than individual pyrrole and pyrimidine nucleus. Different pathological targets require different structural attributes reflected via varied substitutions, thus in recent years, researchers have employed various synthetic strategies to achieve desired substitutions on the pyrrolopyrimidine nucleus. In this review, authors highlight the recent advancement in this area, special focus was laid on the pharmacological profile and structure-activity relationship studies (SAR) of various synthesized pyrrolopyrimidine derivatives.

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## 1. Introduction

Fused pyrimidines are one of the interesting class of heterocycles, which have been extensively explored by medicinal chemists due to their vast pharmacological profile [1]. One of the reasons for such interesting pharmacological potential could be the presence of fused pyrimidines in several physiological molecules. Several fused pyrimidine molecules such as purines, xanthines, pteridines, alloazines, quinazolines, pyrrolopyrimidines, pyridopyrimidines, triazolo pyrimidines, pyrimidoazepines, furopyrimidines and thiazolo pyrimidines are well-established as antibacterial, anticancer, antifungal, and anti-inflammatory agents [1]. Some of these derivatives containing fused pyrimidine nucleus are shown in Fig. 1. Among various fused pyrimidines, pyrrolopyrimidines have gained attention of many researchers as a useful scaffold with varied biological activities such as anticancer, antibacterial, anti-inflammatory and antiviral agents. The initial work on pyrrolopyrimidines began with the discovery of purine derivatives e.g. 6-mercaptopurine as antitumor agent. Initial studies were followed by exhaustive research focussed on pharmacological spectrum of purines and its analogues, in particular deaza analogues, pyrrolo

[2,3-d]pyrimidines and pyrrolo[3,2-d]pyrimidines [2]. Additionally in the mid-nineteenth century, a few natural product based nucleoside antibiotics, tuberidin, toyocamycin, and sanguivomycin were also isolated and identified. Interestingly enough all of these natural product based anti-biotics consisted of pyrrolo[2-3-d]pyrimidine nucleus [2]. Later with the disclosure of different biological activities, especially antitumor potential of substituted 4-aminopyrrolo-[3,2-d]-pyrimidines, researchers shifted their focus towards the second isomer. Some of the FDA approved anti-cancer drugs also contain pyrrolopyrimidine nucleus including Pemetrexed used as antifolate [3], Ruxolitinib, Tofacitinib [4], Baricitinib [5] and Ribociclib [6] used as tyrosine kinase inhibitors, as shown in Fig. 2. Currently, some pyrrolopyrimidine derivatives are in clinical trials against various pathological targets. AZD5363, reported as potent Akt inhibitor, is currently in phase I of clinical trial, in combination with docetaxel and prednisolone for the treatment of metastatic castration resistant prostate cancer (mCRPC). The phase I clinical trial data suggests that in phase II, dose of 320 mg of AZD5363 two times a day and in combination with docetaxel and prednisolone (full dose) is recommended for continuous 4 days [7]. Additionally, due significant preclinical results of AZD5363 in combination with enzalutamide (ENZ) against castrate-resistant prostate cancer in ENZ-resistant cell line, this combination has been put forward for clinical evaluation [8]. TAK-285, a tyrosine

\* Corresponding author.  
E-mail address: [raval.ravindra@gmail.com](mailto:raval.ravindra@gmail.com) (R.K. Rawal).

## APPENDIX

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Review article

**Role of sulphur-heterocycles in medicinal chemistry: An update**

Shelly Pathania <sup>a,b</sup>, Raj Kumar Narang <sup>a</sup>, Ravindra K. Rawal <sup>c,\*</sup>

<sup>a</sup> Department of Pharmaceutical Chemistry, Indo-Soviet Friendship College of Pharmacy (ISCP), Moga, 142001, Punjab, India  
<sup>b</sup> Research Scholar, Maharaja Ranjit Singh Punjab Technical University, Bathinda, Punjab, 151001, India  
<sup>c</sup> Department of Chemistry, Maharishi Markandeshwar (Deemed to Be University), Mullana, 133207, Haryana, India

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**ABSTRACT**

From many decades, S-heterocycles have maintained their status as an important part and core of FDA approved drugs and medicinally active compounds. With exhaustive exploration of nitrogen heterocycles in medicinal chemistry, researchers have shifted their interest towards other heterocycles, especially, S-heterocycles. Thus several attempts have been made to synthesize a variety of new sulphur containing compounds with high medicinal value and low toxicity profile, in comparison to previous N-heterocycles. Till today, S-heterocycle containing compounds have been largely reported as anticancer, antidiabetic, antimicrobial, antihypertension, antiviral, antiinflammatory etc. In this review, the authors have tried to provide a critical analysis of synthesis and medicinal attributes of sulphur containing heterocycles such as thiirane, thiophene, thiazole, thiopyran, thiazolidine etc reported within last five years to emphasize the significance and usefulness of these S-heterocycles in the drug discovery process.

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\* Corresponding author.  
E-mail address: rawalravindra@gmail.com (R.K. Rawal).

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## Review

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## An update on chemical classes targeting ERK1/2 for the management of cancer

Shelly Pathania<sup>1,2</sup> & Ravindra K Rawal<sup>1,\*</sup> <sup>1</sup>Department of Pharmaceutical Chemistry, Indo Soviet Friendship College of Pharmacy, Moga 142001, Punjab, India<sup>2</sup>Research Scholar, Maharaja Ranjit Singh Punjab Technical University, Bathinda 151001, Punjab, India<sup>3</sup>Department of Chemistry, Maharsi Markandeshwar (deemed to be University), Mullana 133207, Ambala, Haryana, India

\*Author for correspondence: Tel: +91 1731 304 229; rawal.ravindra@gmail.com

Cancer, still in the limelight due to its dreadful nature, shows overexpression of multiple signaling macromolecules leading to failure of many chemotherapeutic agents and acquired resistance to chemotherapy. These factors highlight the significance of shifting toward targeted therapy in cancer research. Recently, ERKs (ERK1 and 2) have been established as a promising target for the management of various types of solid tumors, due to their aberrant involvement in cell growth and progression. Several ERKs inhibitors have reached clinical trials for the management of cancer and their derivatives are being continuously reported with noteworthy anticancer effect. This review highlights the recent reports on various chemical classes involved in the development of ERKs inhibitors along with their *in vitro* and *in vivo* activity and structure-activity relationship profile.

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**Keywords:** cancer • ERK1/ERK2 • ERK inhibitors • heterocycles

From many decades, cancer remains a terrible disease and a challenge for the researchers due to its poor prognosis [1–3]. Although a number of chemotherapeutic agents are available in the market for the treatment of cancer, they fail to cure the disease for various reasons [4,5]. One of the main reasons for their failure is the occurrence of drug resistance. Resistance can occur due to alterations in the genes during the treatment, overexpression of the molecular targets and cross linking of the various signaling pathways [6]. Thus, the need of the hour is to focus on the targeted therapy rather than chemotherapy. Continuously, researchers are putting their efforts into identifying new targeted therapies with great efficacy and less toxicity in comparison to chemotherapy [7,8]. The use of various modern techniques like high-throughput screening (HTS), bioinformatics, genomics and molecular biology helps in the identification of new therapeutic targets with higher degree of relevance in the treatment of cancer [9,10]. Various studies have identified many potential targets and ERKs [11], ERK1 and ERK2 are one of the promising targets among them, which seek the attention of many researchers. ERK1 and ERK2 belong to the mitogen-activated family, thus also known as mitogen-activated protein kinases (MAPKs) [12]. Both ERK1 and ERK2 contain 85% of the same amino acid sequence. These are categorized as RAS–RAF–MEK–ERK signaling cascade due to their active participation in the cell functions like cell growth, cell differentiation and proliferation [13–15]. The proto-oncogenic drivers like mutation leads to dysregulation and increased kinases activity [16]. Due to this increased phosphorylation, ERKs have been involved in various types of cancers like breast, lung, prostate and ovarian cancer [17]. During normal physiological conditions, MAPKs activated by Ras/Raf proteins such as A-Raf, B-Raf or C-Raf and various other growth factors, bind to the surface of the receptors [18]. The activated MAPKs further phosphorylates other downstream isoforms of ERK, especially ERK1 and ERK2, the key kinase effector of the pathway, which further leads to the activation of many transcription factors (Figure 1) [19]. In cancer, there is an abnormal activation of the ERK pathway caused by the overactivation of growth factor receptors and mutations in RAS and BRAF proteins [20]. Approximately 33 and 8% of the human cancers are found to have overexpressed RAS and BRAF proteins, respectively [21]. Thus, researchers get encouraged by these findings and are involved in the development of small-based molecules targeting ERK signaling cascade for the management of cancer. Till date, several ERK inhibitors have been reported; based on their mechanism, ERK inhibitors have been categorized

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## APPENDIX

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### Identifying novel putative ERK1/2 inhibitors via hybrid scaffold hopping–FBDD approach

Shelly Pathania<sup>a,b</sup>, Pankaj Kumar Singh<sup>d</sup>, Raj Kumar Narang<sup>a</sup> and Ravindra K. Rawal<sup>c</sup>

<sup>a</sup>Department of Pharmaceutical Chemistry, ISF College of Pharmacy, Moga, Punjab, India; <sup>b</sup>Maharaja Ranjit Singh Punjab Technical University, Bathinda, Punjab, India; <sup>c</sup>Department of Chemistry, Maharishi Markandeshwar (Deemed to be University), Mullana, Haryana, India; <sup>d</sup>Integrative Physiology and Pharmacology, Institute of Biomedicine, Faculty of Medicine, University of Turku, Turku, Finland

Communicated by Ramaswamy H. Sarma

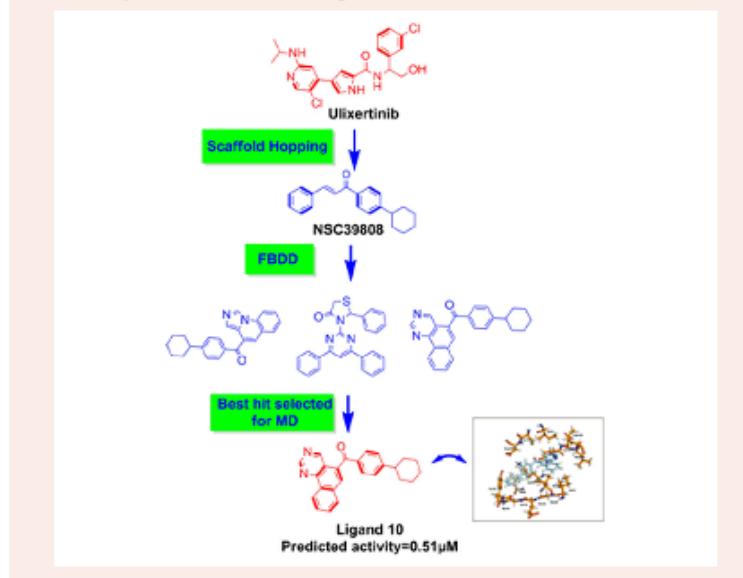
#### ABSTRACT

ERK inhibitors are continuously explored by the researchers due to their clinical significance in resistant tumor cell lines. Though many ERK1/2 inhibitors are reported, there is still need to identify novel hits to increase the number of molecules in clinical trials. Therefore, an urgent need is to examine the existing chemical space for ERK inhibitory potential with an aim to develop novel scaffolds which can act as potent ERKs inhibitors. In the study, Ulixertinib, a known ERK2 inhibitor was selected to perform scaffold hopping to discover new scaffolds with similar binding mode followed by molecular docking analysis of the hits with highest similarity score to determine, both the binding mode and affinity in the catalytic domain of ERK2. The top hit was then subjected to FBDD to identify side chains which could enhance the binding affinity in the catalytic domain of ERK2. Again, docking analysis was performed to validate and determine their binding affinity. Further the top hit identified after docking analysis was subjected to molecular dynamic simulations. Overall, 3 hits (Ligand 6, 8 and 10) were found to possess optimum pharmacodynamic and pharmacokinetic profile, *in-silico*, to be claimed as putative ERK2 inhibitors. This study disclosed new lead molecules with putative ERK2 inhibitory potential which may be further validated via biological evaluation.

#### ARTICLE HISTORY

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**KEYWORDS**  
ERK2 inhibitors; *in-silico*; Scaffold hopping; FBDD; molecular docking



**CONTACT** Ravindra K. Rawal [ravala@vindra@gmail.com](mailto:ravala@vindra@gmail.com) Department of Chemistry, Maharishi Markandeshwar (Deemed to be University), Mullana, Haryana 133207, India  
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## APPENDIX

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### Structure based designing of thiazolidinone-pyrimidine derivatives as ERK2 inhibitors: Synthesis and in vitro evaluation

S. Pathania<sup>a,b</sup>, P.K. Singh<sup>c</sup>, R.K. Narang<sup>a</sup> and R.K. Rawal<sup>d,e</sup>

<sup>a</sup>Department of Pharmaceutical Chemistry, ISF College of Pharmacy, Moga, India; <sup>b</sup>Department of Pharmaceutical Sciences & Technology, Maharaja Ranjit Singh Punjab Technical University, Bathinda, India; <sup>c</sup>Integrative Physiology and Pharmacology, Institute of Biomedicine, Faculty of Medicine, University of Turku, Turku, Finland; <sup>d</sup>Department of Chemistry, Mahanshi Markandeshwar (Deemed to Be University), Ambala, India; <sup>e</sup>CSIR-North East Institute of Science and Technology, Jorhat, India

#### ABSTRACT

Breast cancer has been associated with an overexpression of various molecular targets; accordingly, various target-specific chemotherapeutic agents have been developed. Inhibition of ERK2, a member of MAPK pathway, is an important target involved in the treatment of both oestrogen receptor-positive and triple-negative breast cancer. Thus, in continuation of our previous work on the ERK2 target, we here report novel inhibitors of this kinase. Out of three lead molecules reported in our previous study, we selected the thiazolidinone-pyrimidine scaffold for further development of small molecule inhibitors of ERK2. Analogue of the lead molecule were docked in the target kinase, followed by molecular dynamic simulations and MM-GBSA calculations. Analogue maintaining key interactions with amino acid residues in the ATP-binding domain of ERK2 were selected and duly synthesized. In vitro biochemical evaluation of these molecules against ERK2 kinase disclosed that two molecules possess significant kinase inhibitory potential with IC<sub>50</sub> values ≤ 0.5 μM.

#### ARTICLE HISTORY

Received 10 July 2021  
Accepted 23 August 2021

#### KEYWORDS

Thiazolidinone-pyrimidine;  
molecular dynamic  
simulations; ERK2; enzymatic  
assay

#### Introduction

Breast cancer forms the second leading cause of death around the globe and has an adverse impact on women's health [1]. On the basis of overexpression of molecular targets, breast cancer is classified into many types such as ER/PR+ (overexpression of oestrogen and progesterone receptors; 70% of all breast cancers) and HER2+ (human epidermal growth factor receptor 2; 20% of breast cancers) [2]. A critical subtype of breast cancer is triple-negative breast cancer (TNBC), which is the most aggressive form of breast cancer and considered as most lethal because of the absence of molecular targets such as HER2, ER and PR [2]. As TNBC lacks overexpression of ER, PR and HER2, it is resistant towards hormonal and targeted therapies. However, pathways like mitogen-activated protein kinase (MAPK) have been reported to be

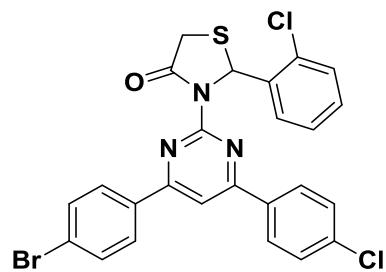
CONTACT R.K. Rawal rawal.ravindra@gmail.com

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# *Appendix*

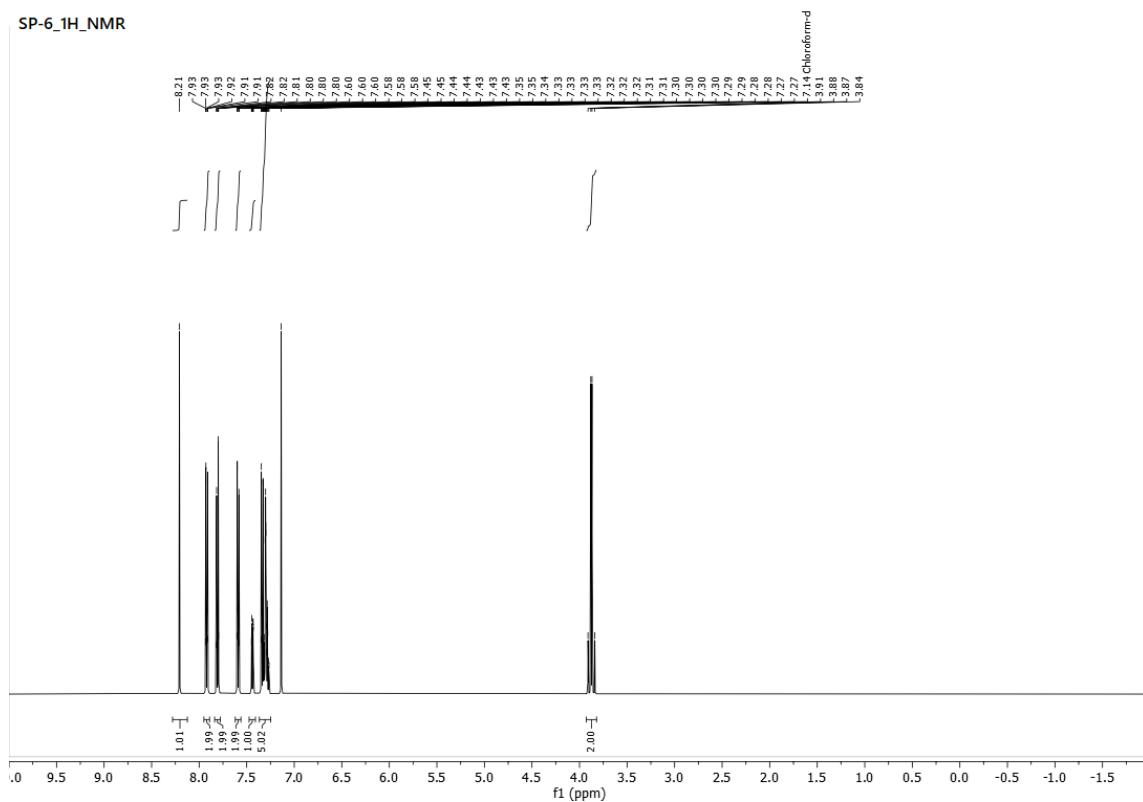
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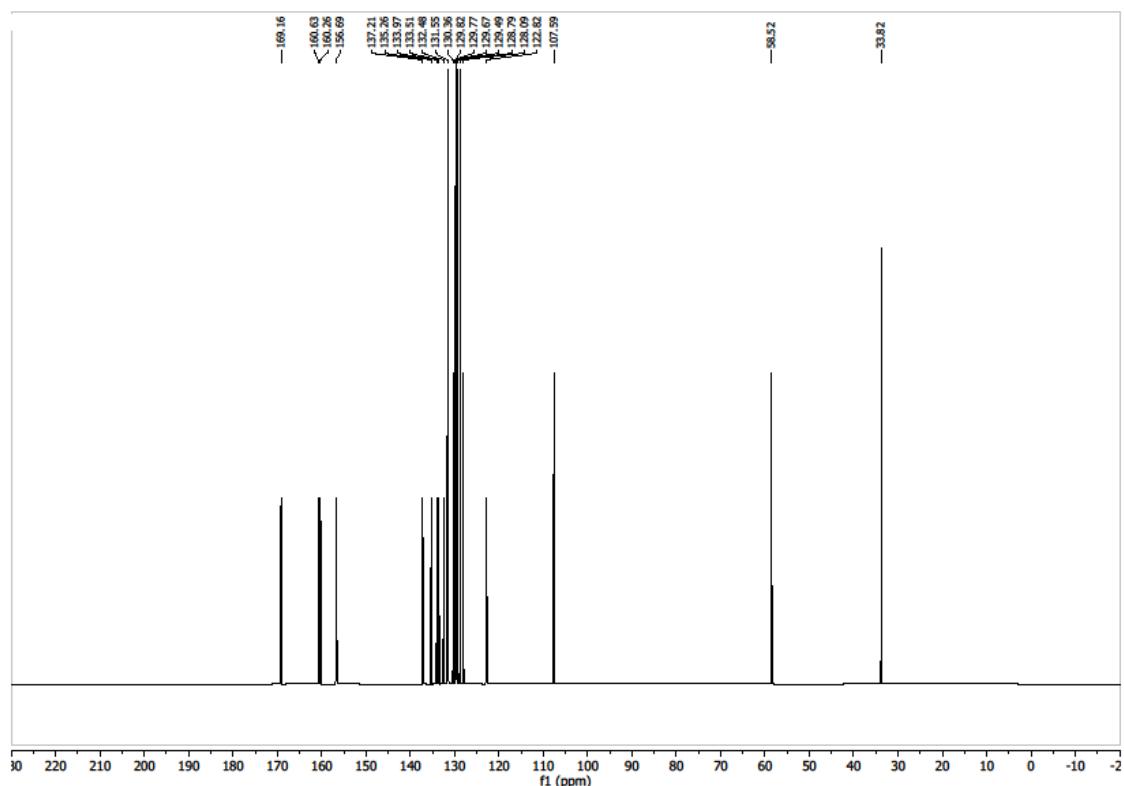
### **<sup>1</sup>H NMR:**

SP-6\_1H\_NMR



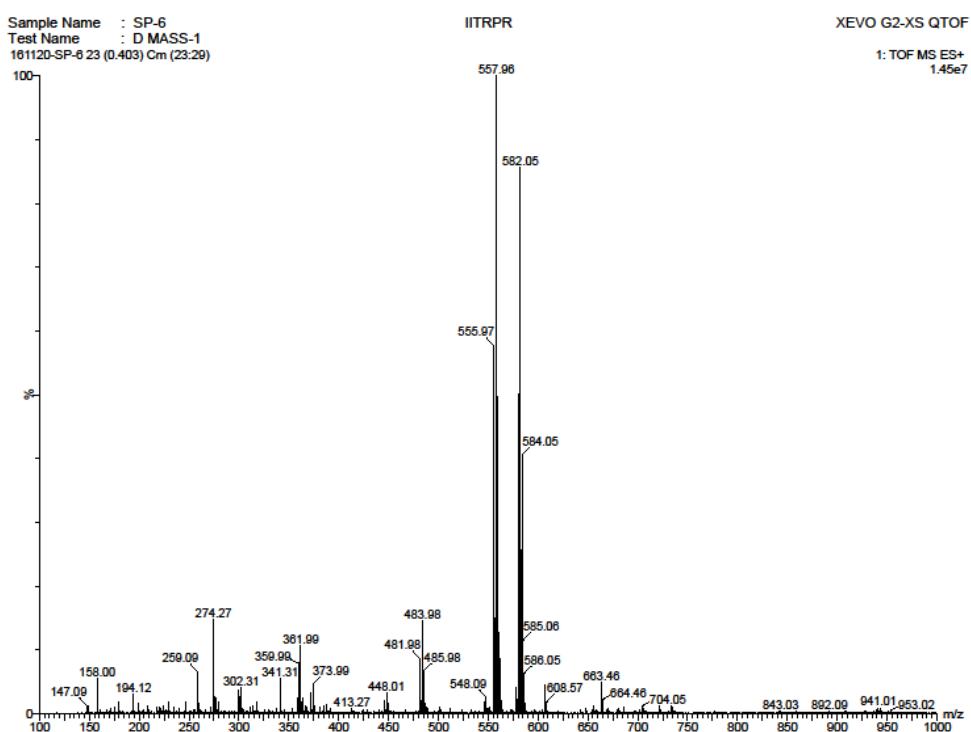
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### <sup>13</sup>C NMR:



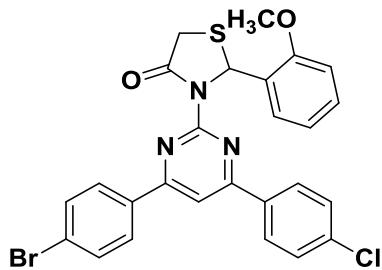
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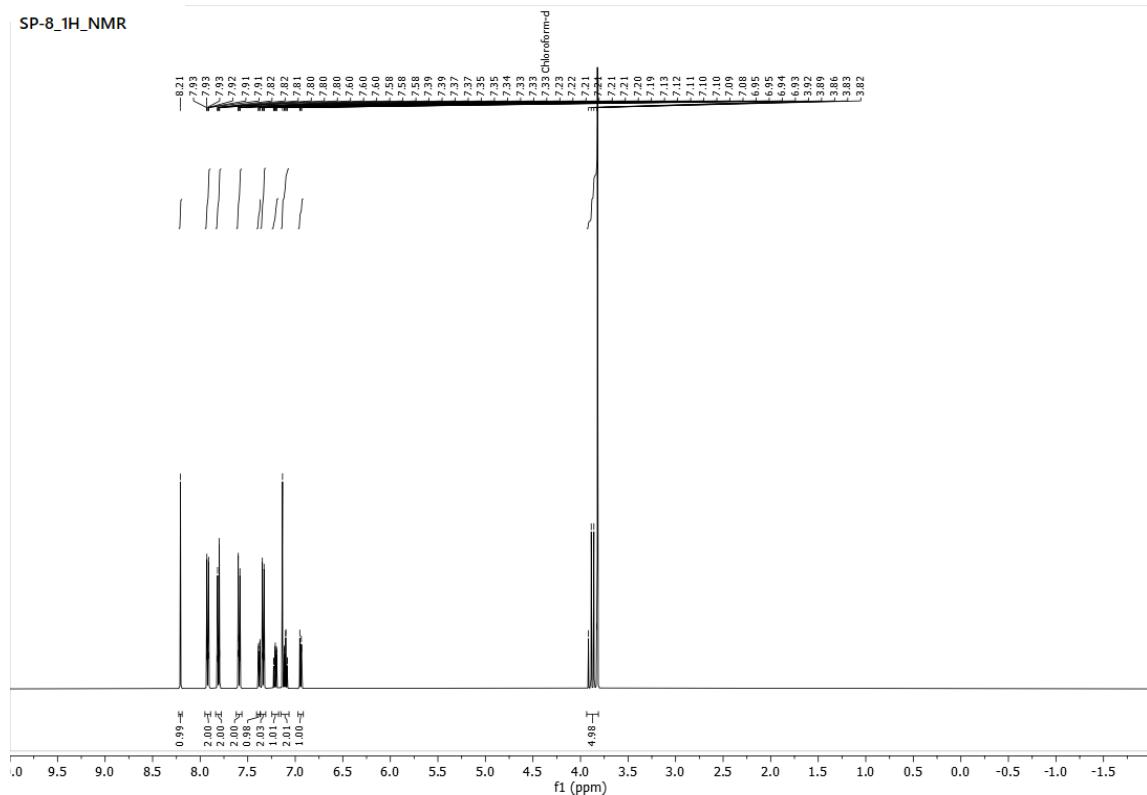


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## Compound 8c:

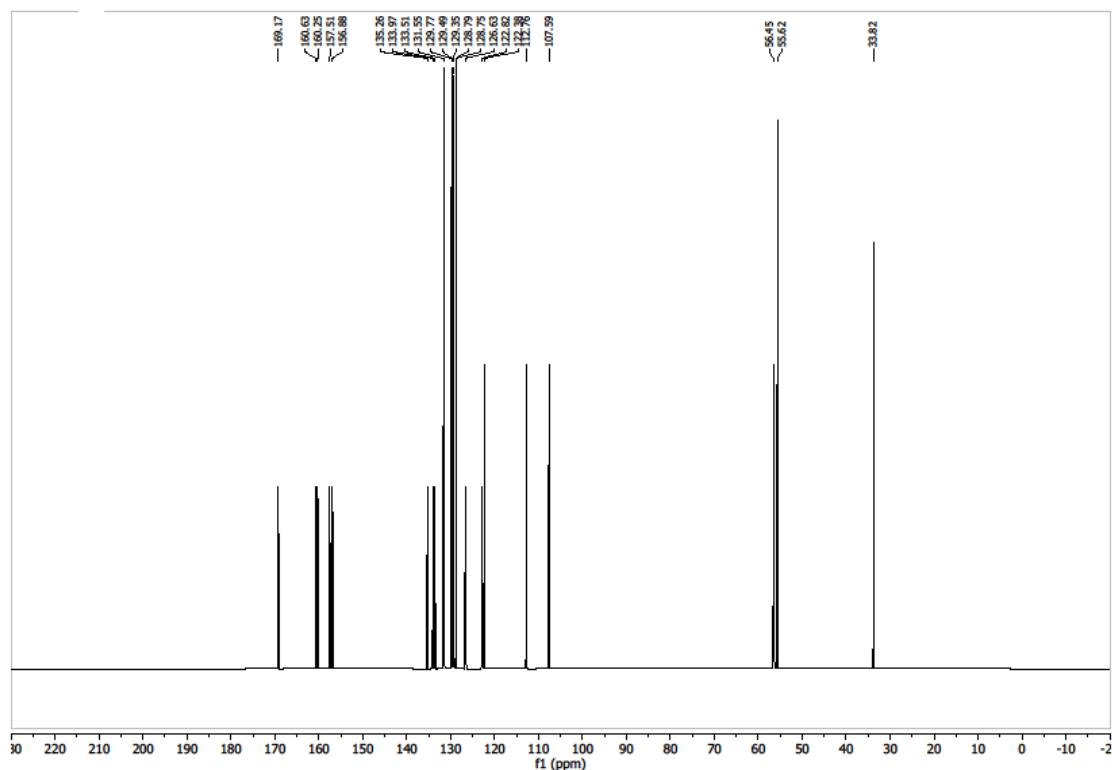


### **<sup>1</sup>H NMR:**



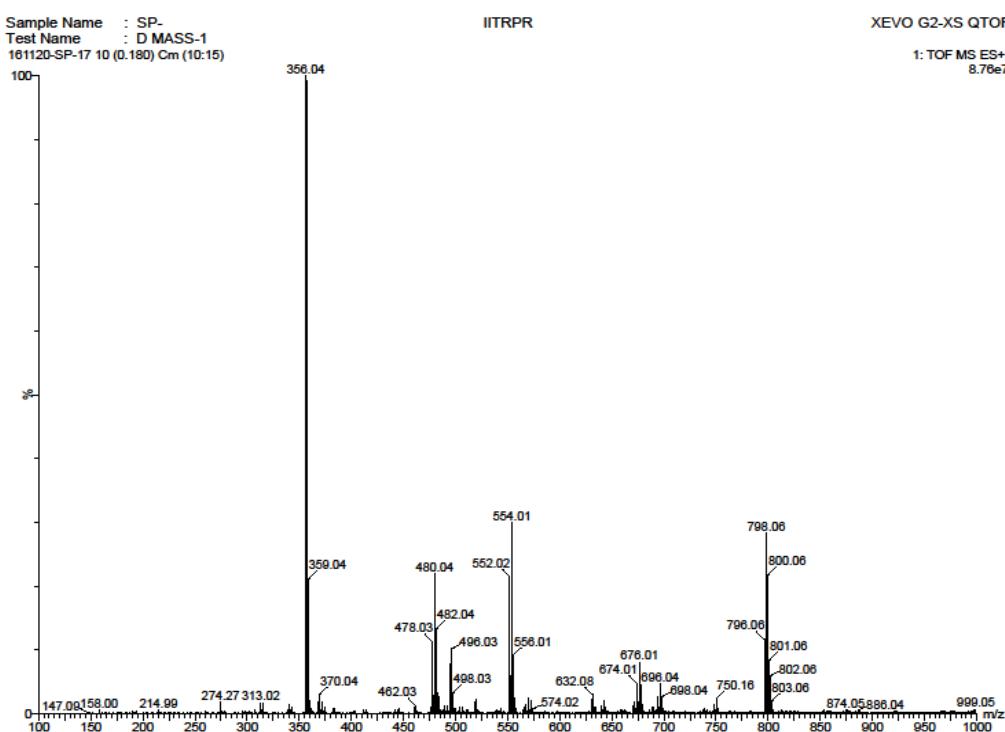
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### <sup>13</sup>C NMR:



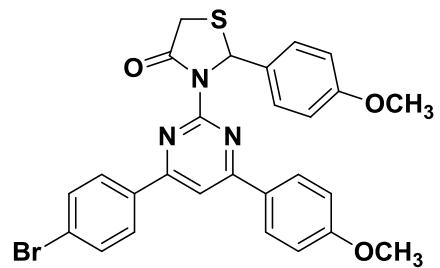
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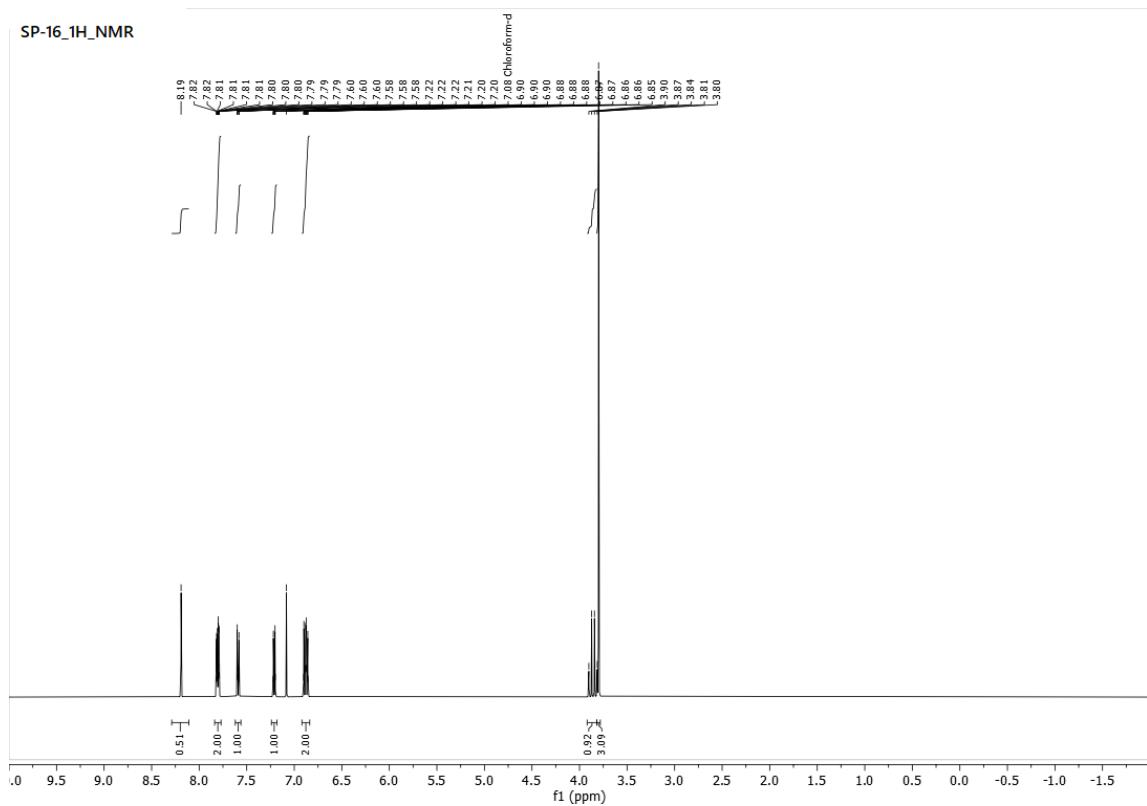


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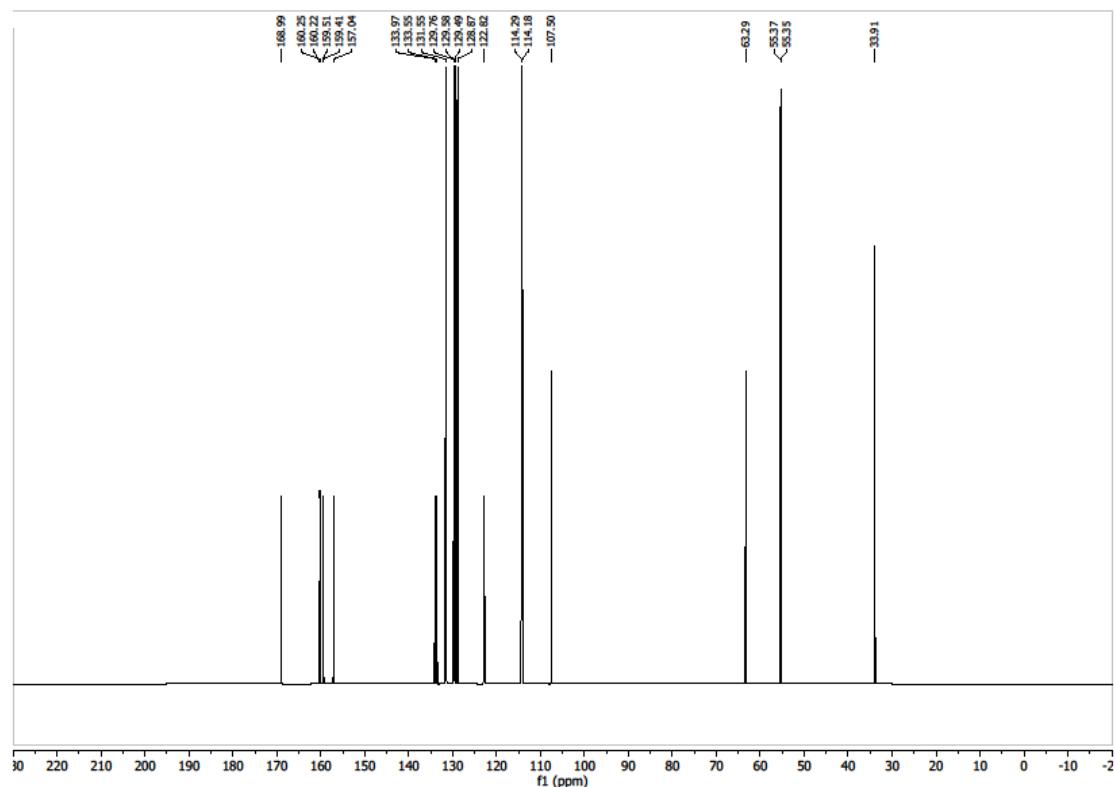


### $^1\text{H}$ NMR:



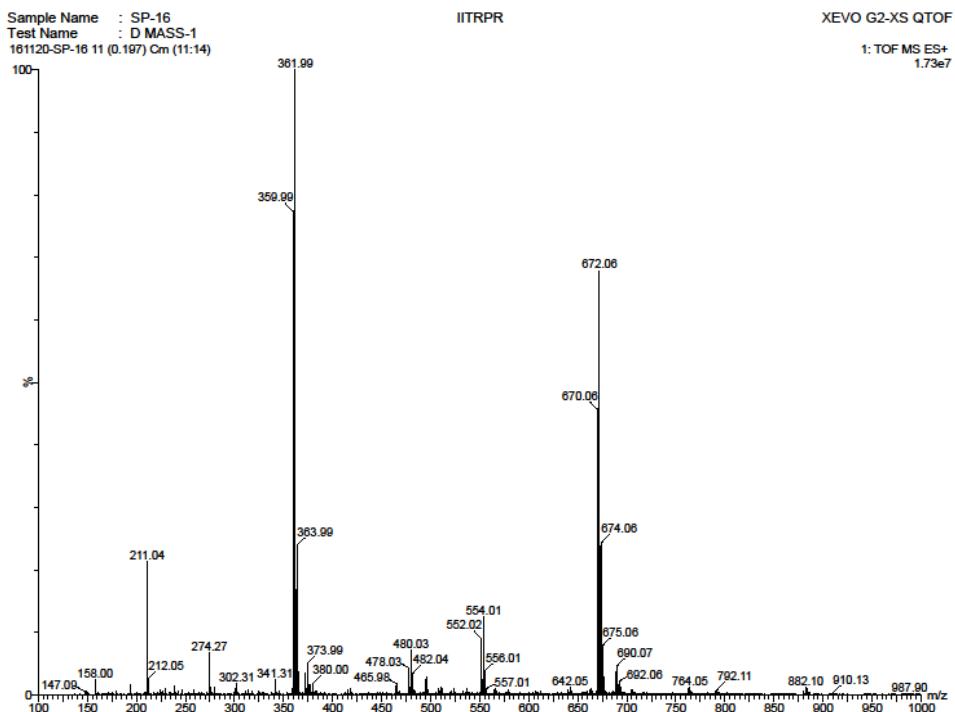
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### <sup>13</sup>C NMR:



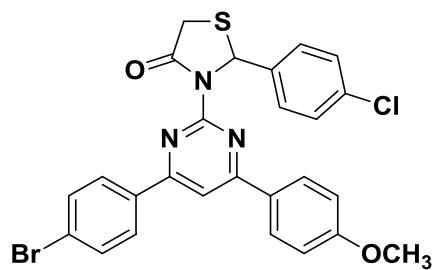
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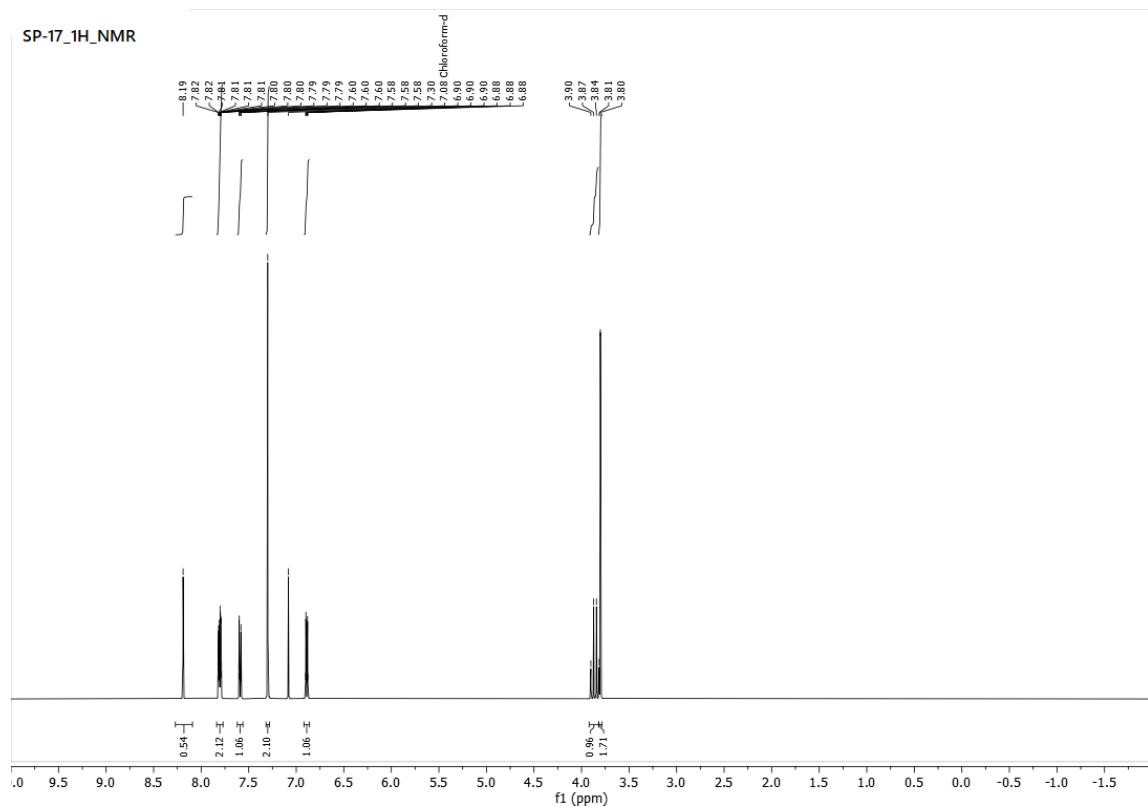


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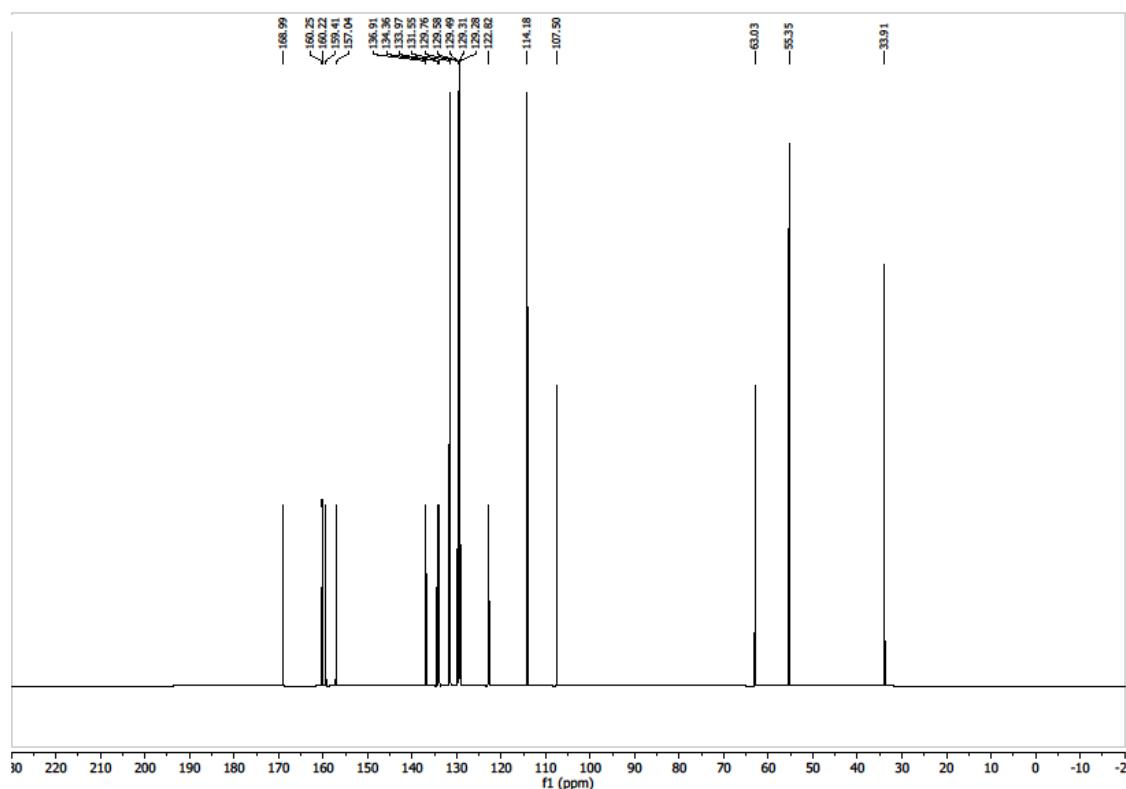


### $^1\text{H}$ NMR:



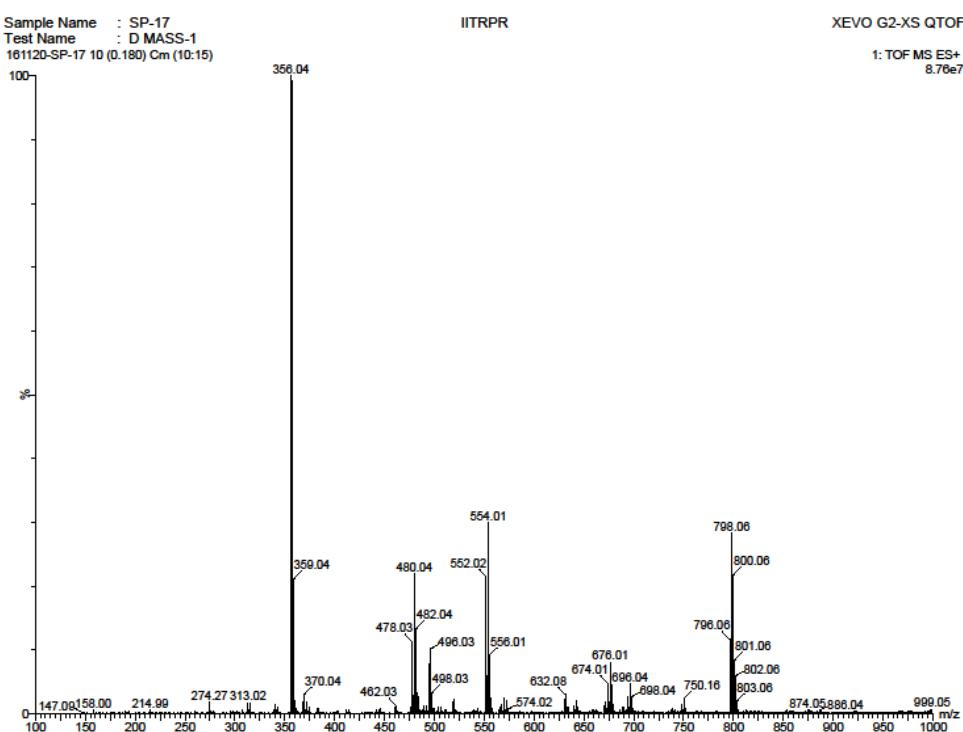
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### <sup>13</sup>C NMR:



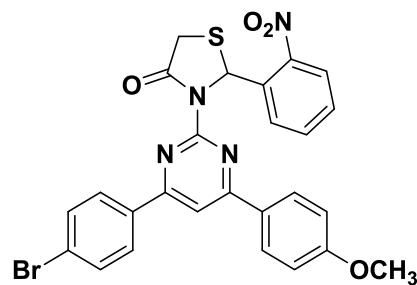
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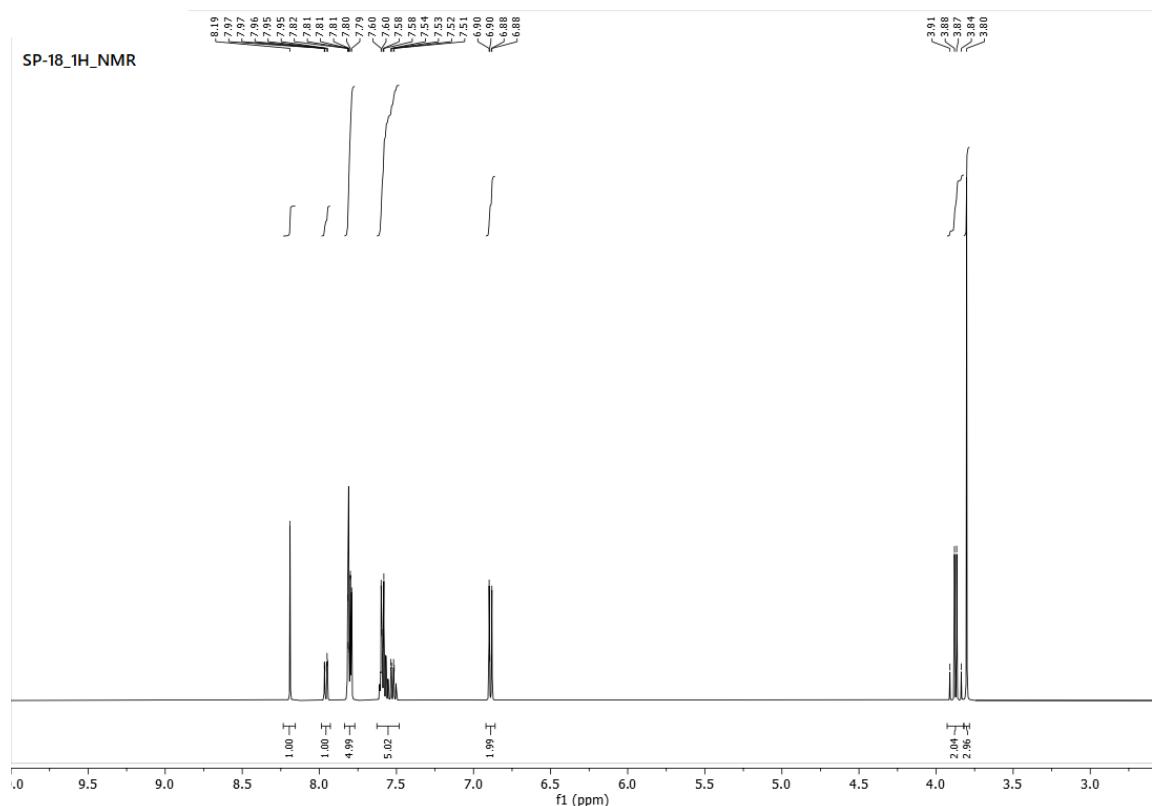


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### **Compound 8l:**

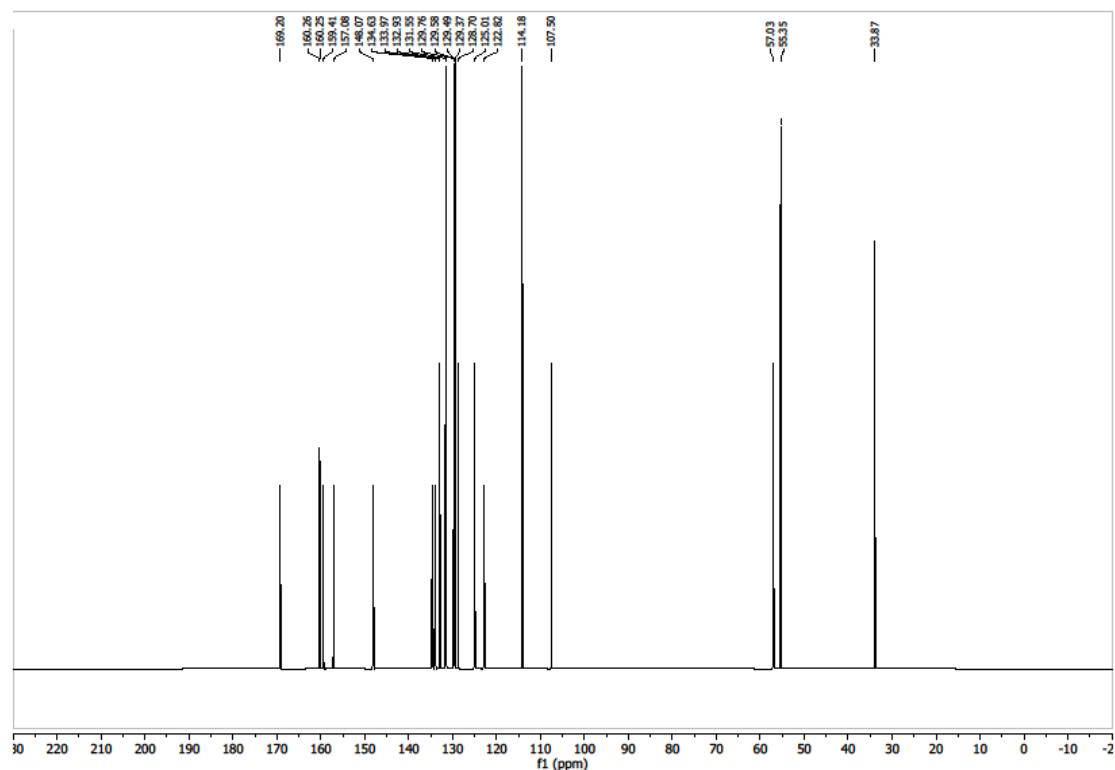


### **<sup>1</sup>H NMR:**



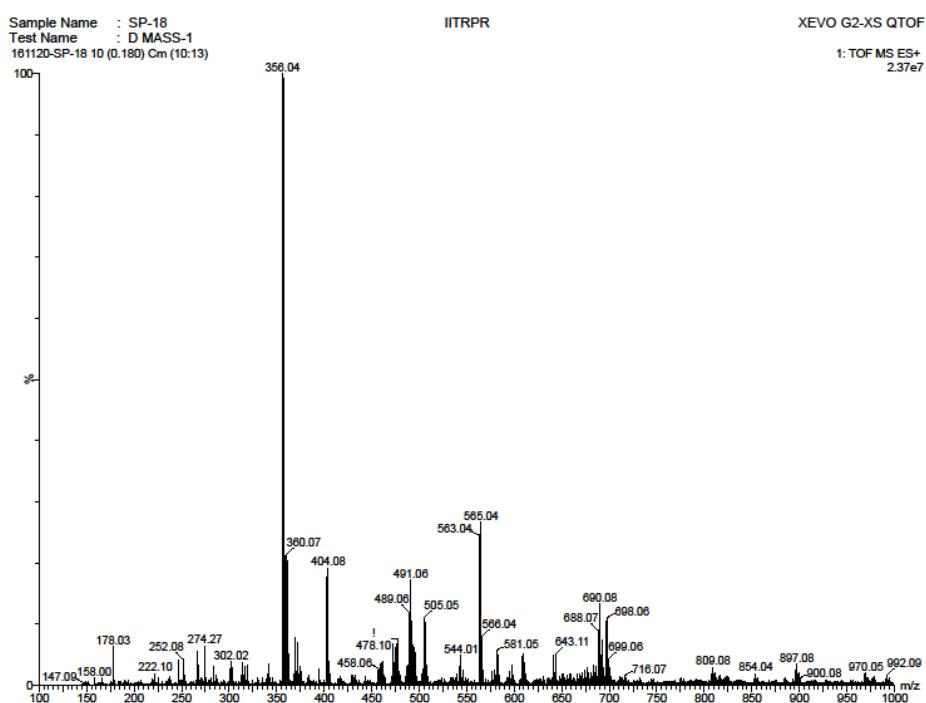
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### <sup>13</sup>C NMR:



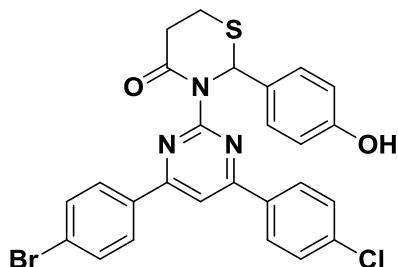
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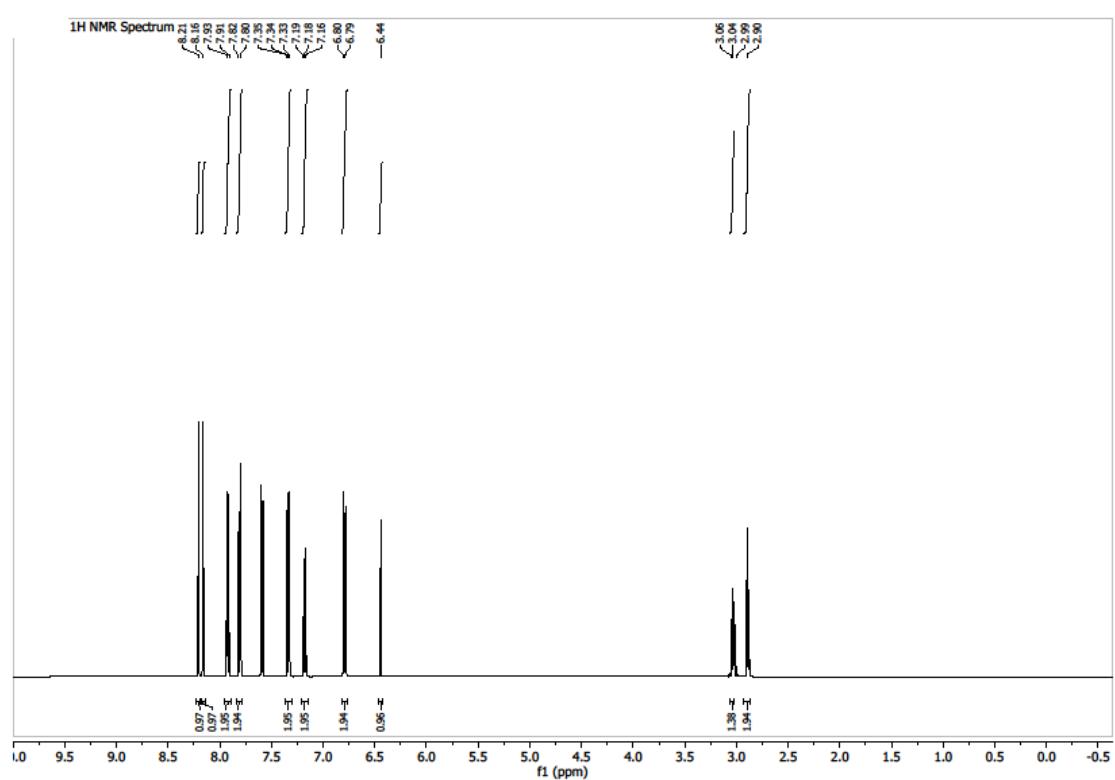


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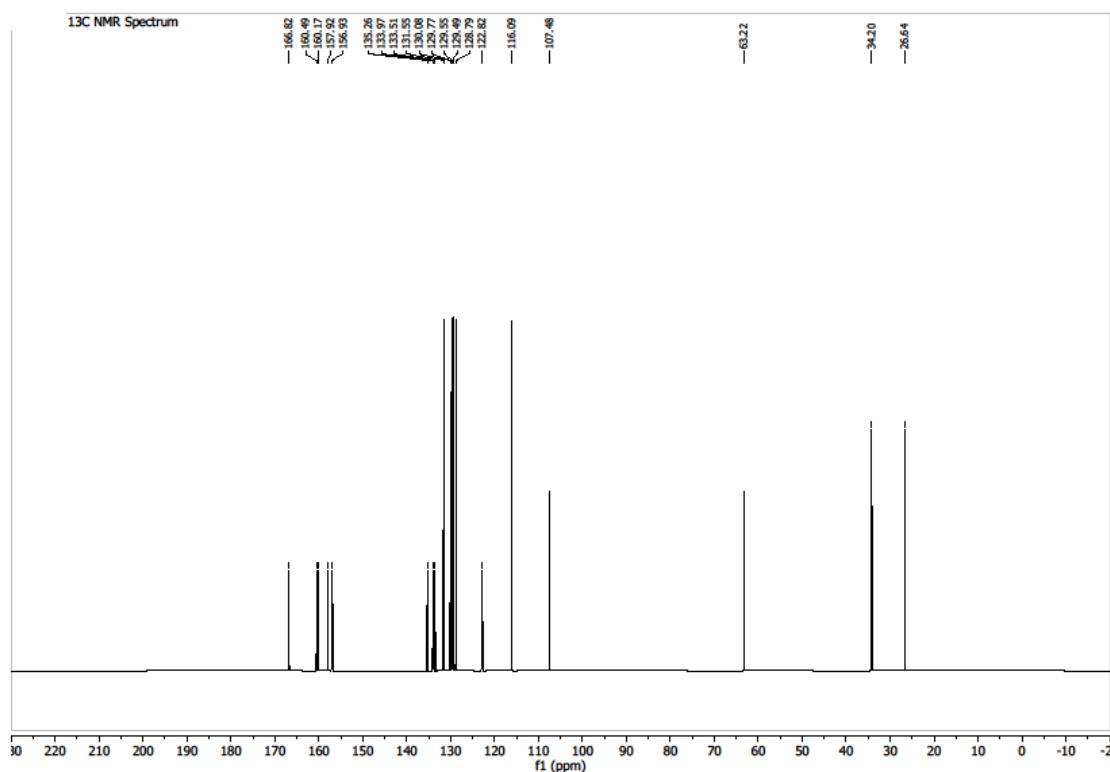


### $^1\text{H}$ NMR:



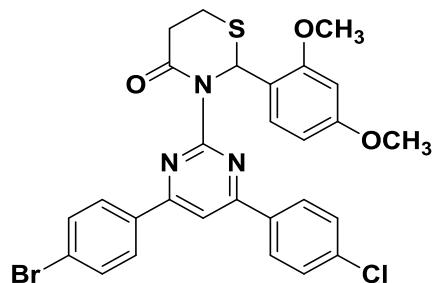
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### <sup>13</sup>C NMR:

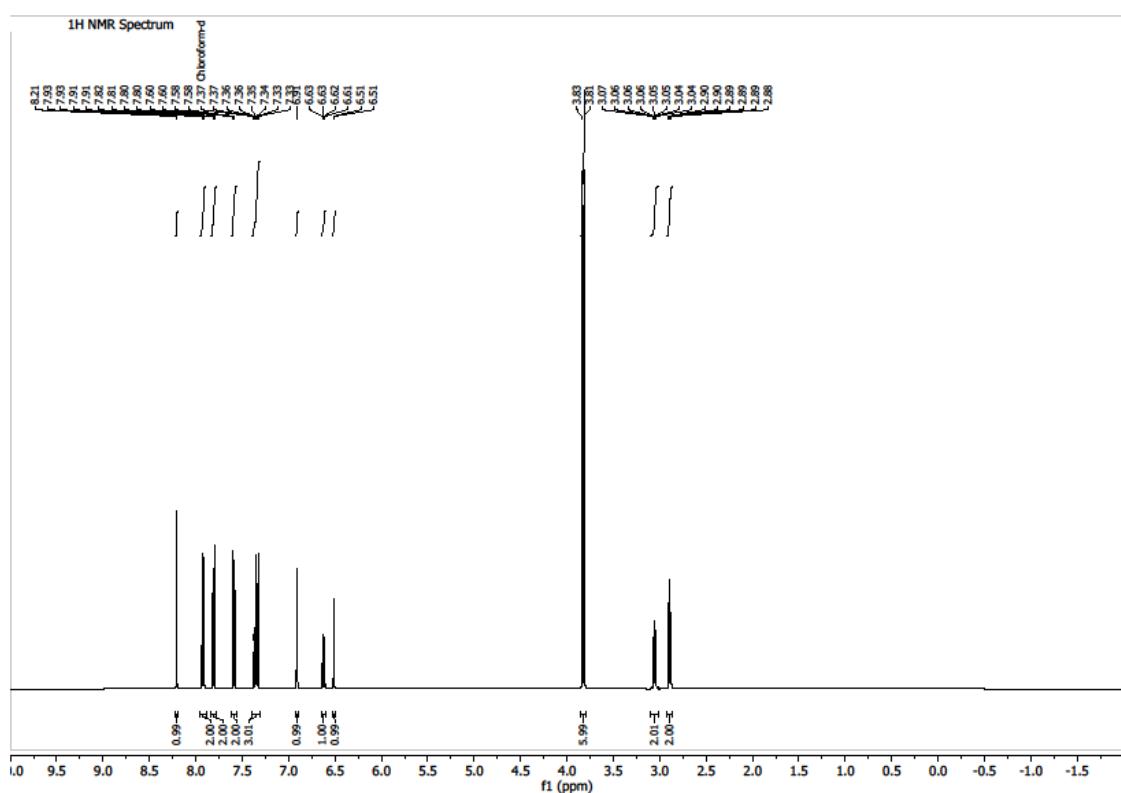


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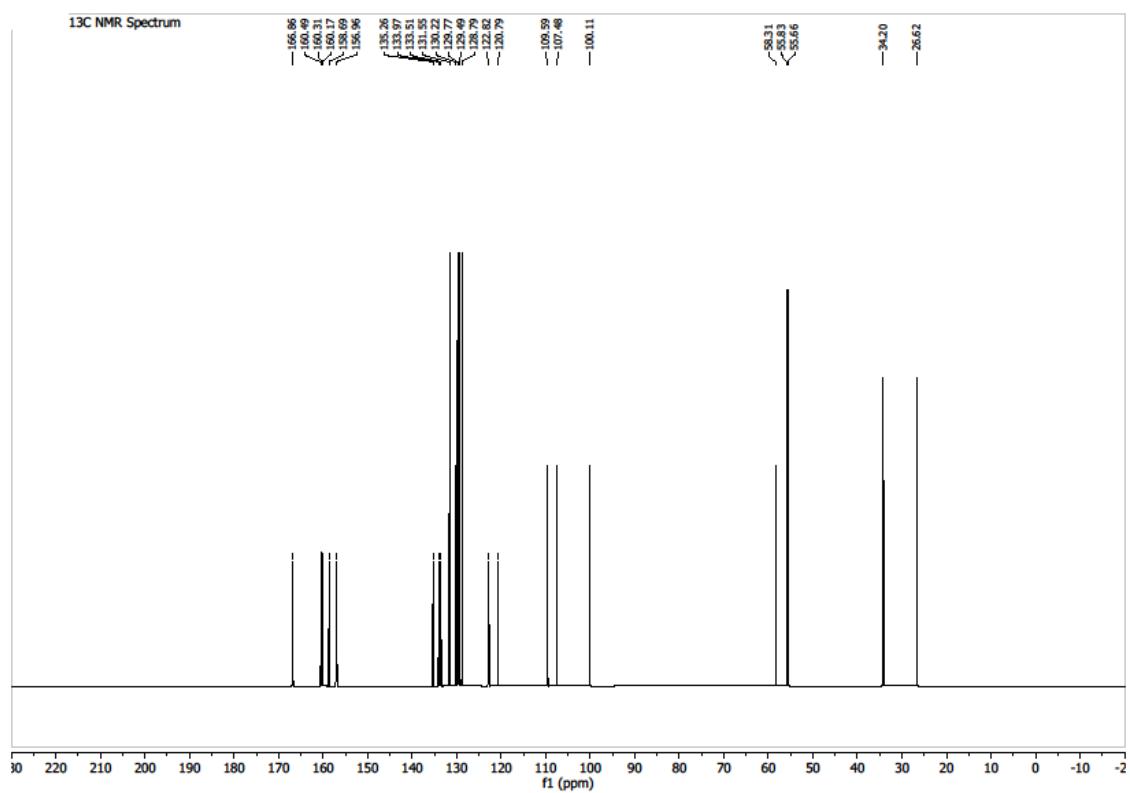


### $^1\text{H}$ NMR:



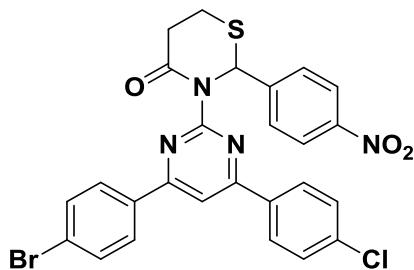
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### <sup>13</sup>C NMR:

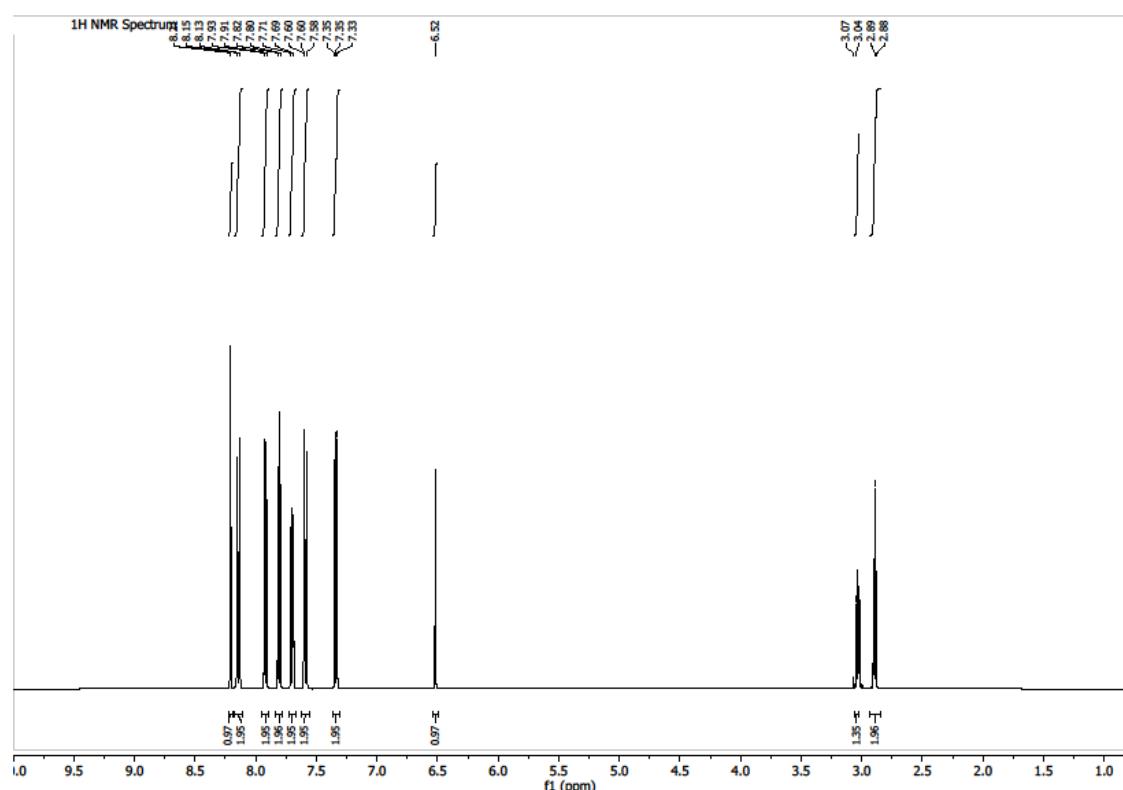


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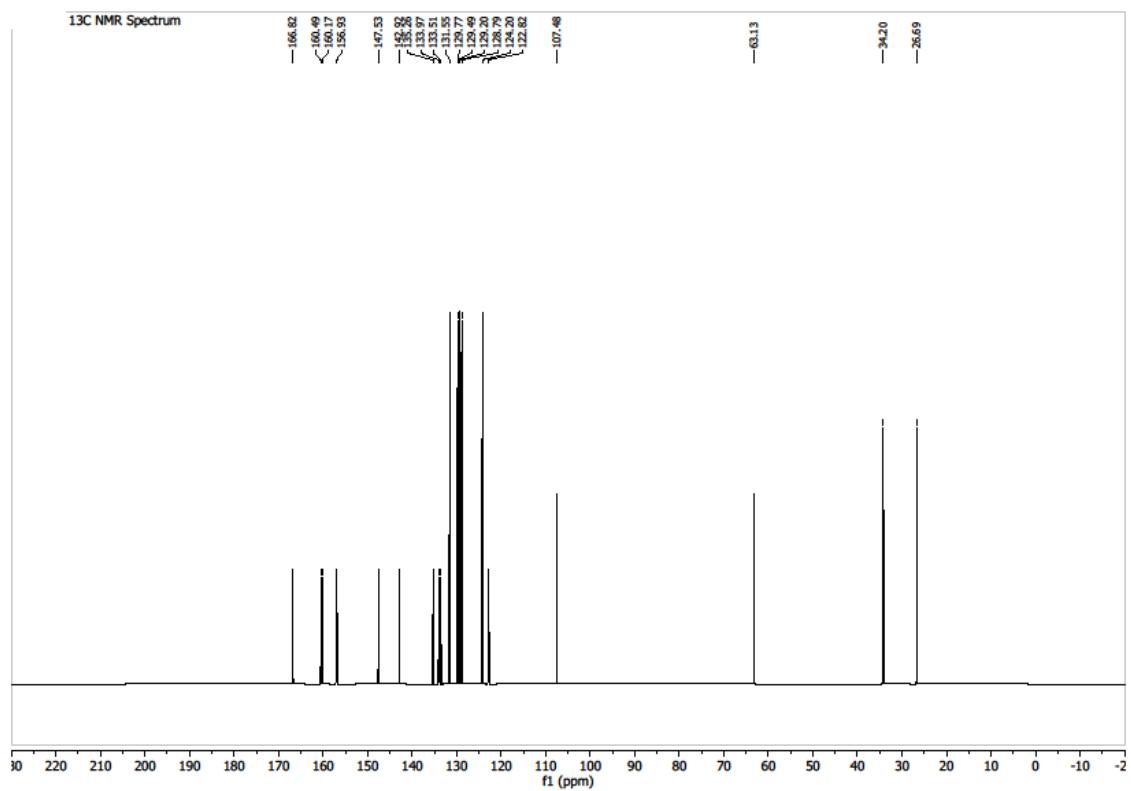


### <sup>1</sup>H NMR:



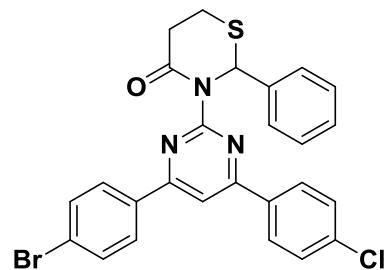
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### <sup>13</sup>C NMR:

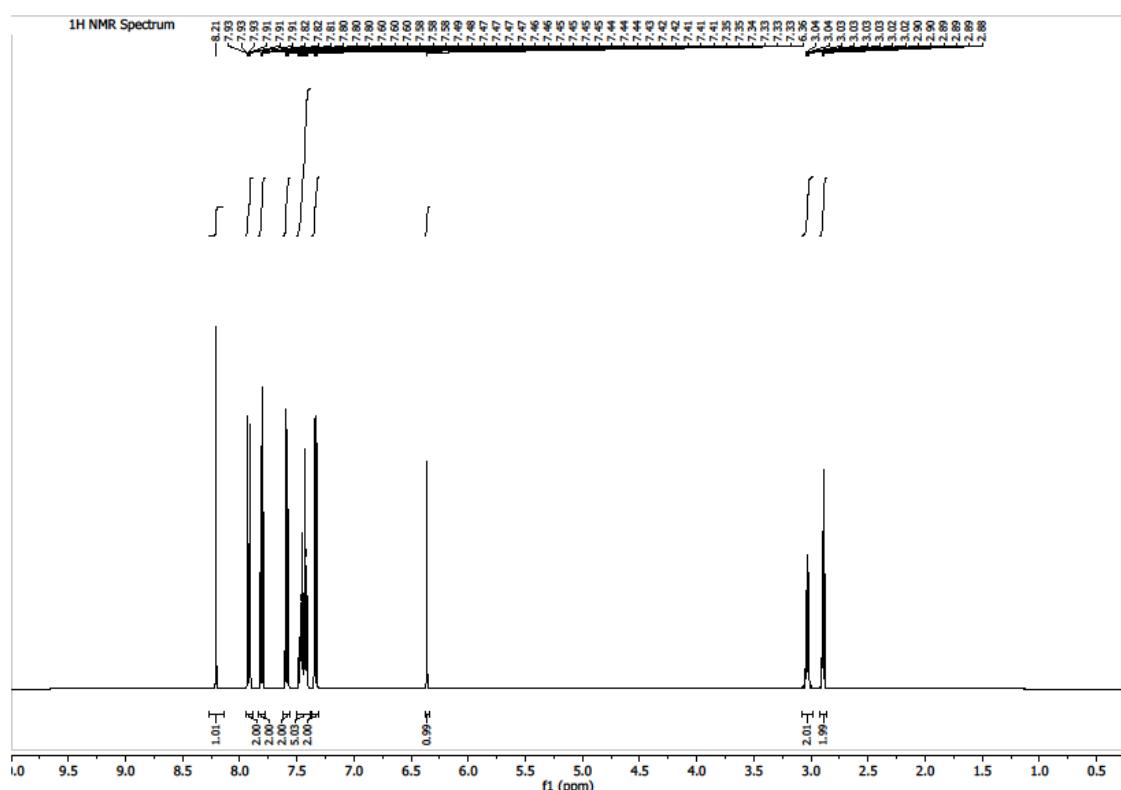


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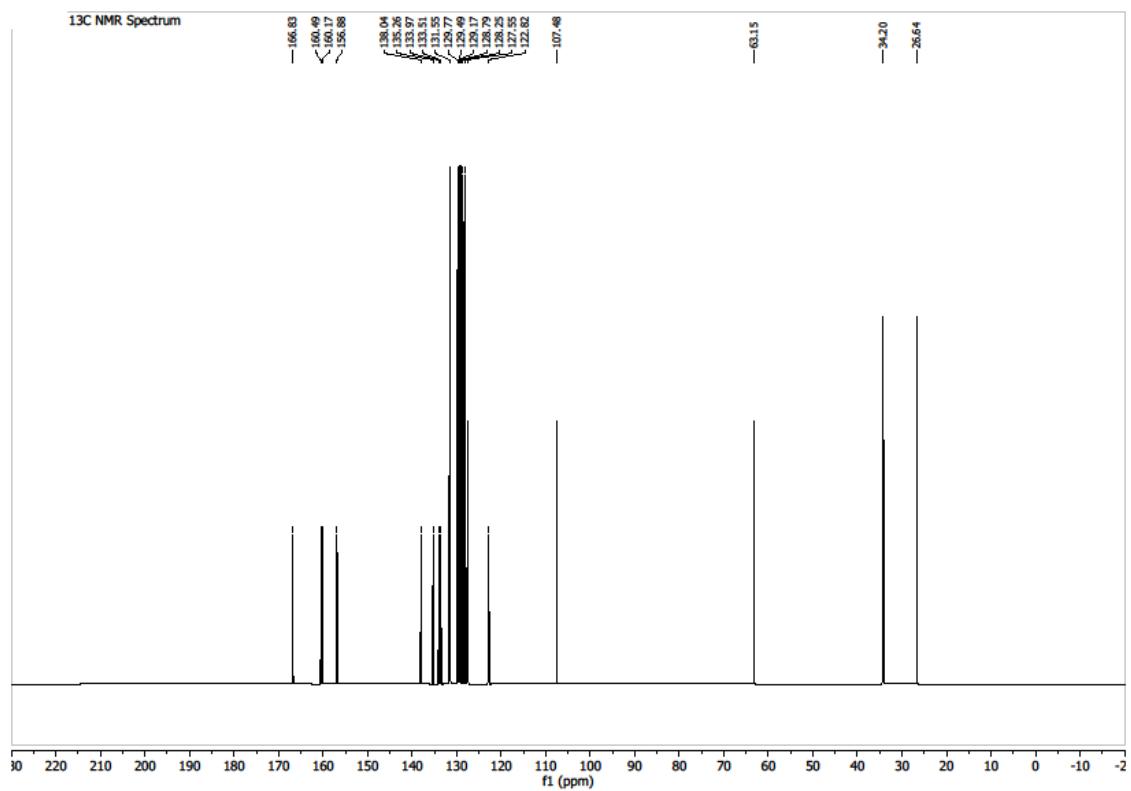


### $^1\text{H}$ NMR:



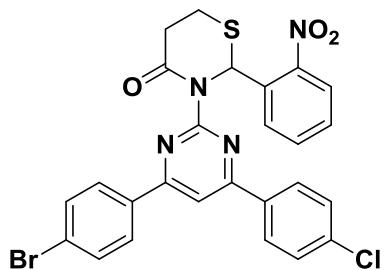
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### <sup>13</sup>C NMR:

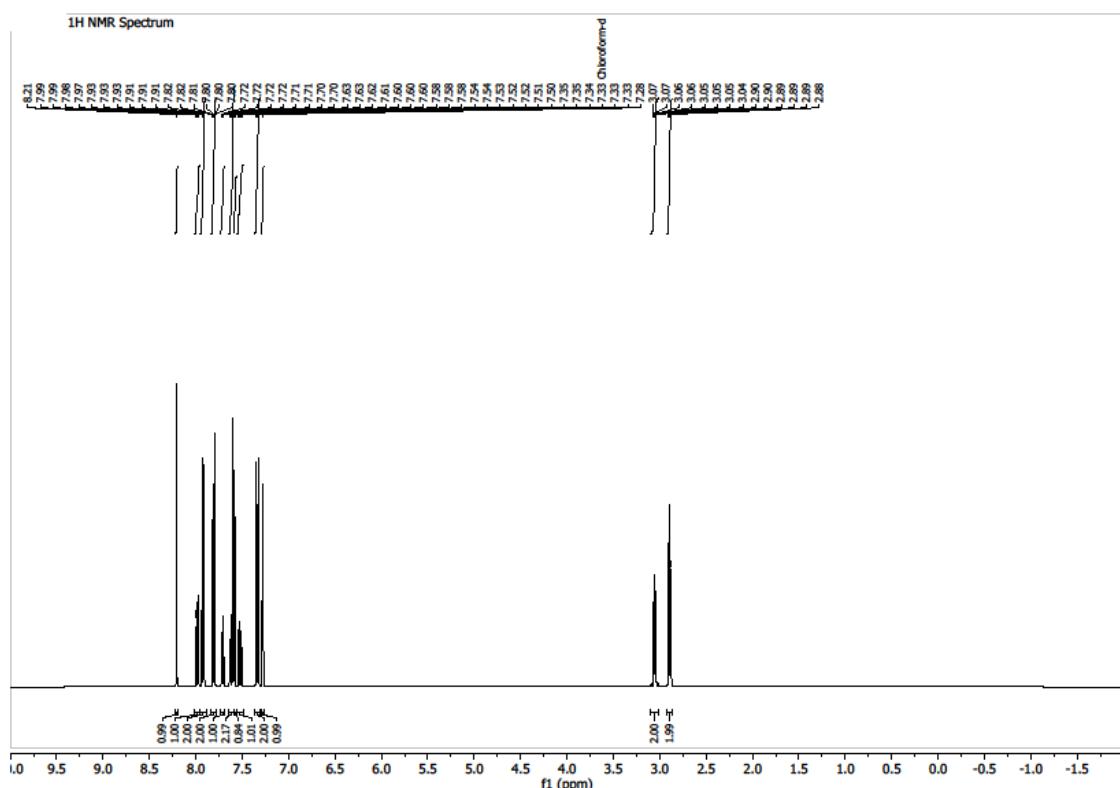


## APPENDIX

### Compound 10i:



### <sup>1</sup>H NMR:



## APPENDIX

### <sup>13</sup>C NMR:

